Amyloplasts and Vacuolar Membrane Dynamics in the Living Graviperceptive Cell of the Arabidopsis Inflorescence Stem

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INTRODUCTION

Higher plants cannot escape from the place where they germinate and settle, even if the environmental conditions drastically change. Plants have developed many mechanisms during the course of evolution to survive in such circumstances by changing growth direction or architecture of their body shape. Typical of such growth responses, gravitropism is one of most prominent phenomena and has been the focus of studies on many plant species over a long period of time (Knight, 1806; Sack, 1991; Fukaki et al., 1996b; Chen et al., 1999). In higher plants, shoots, roots generally show negative and positive gravitropism, respectively. This response ensures an appropriate positioning of tissues for efficient photosynthesis, gas exchange, and the supply of water and minerals.

The gravitropic response in plants can be divided into four sequential steps: gravity perception, signal formation in the gravity perceptive cell, intracellular and intercellular signal transduction, and an asymmetric cell elongation between the upper and lower sides of the responding organs (Tasaka et al., 1999). Some classical hypotheses have been supported by recent molecular genetical studies. For example, the starch-statolith hypothesis is involved in the first step and proposes that the sedimenting amyloplasts, which contain dense starch granules, act in sensing the direction of gravity (Sack, 1991; Weise and Kiss, 1999). However, the mechanism functioning in gravity perception has not yet been characterized at the molecular level.

A recent molecular approach using Arabidopsis thaliana revealed the graviperceptive cell in an inflorescence stem. We have isolated several shoot gravitropism (sgr) mutants, which are defective in the gravitropic response of inflorescence stems. In the Arabidopsis shoot, the epidermis, cortex, endodermis, and stele containing vascular tissues are arranged in a radially symmetrical manner. In the course of the study of sgr1 and sgr7, which are allelic to scarecrow and short-root, respectively, the endodermis has been revealed as essential for the gravitropism of the inflorescence stem (Fukaki et al., 1998; Tasaka et al., 1999). In addition, it is known that amyloplasts differentiate and sediment to the direction of gravity in the endodermis. These findings suggest that the endodermis in the inflorescence stem is essential for the first step of shoot gravitropism, namely the perception of gravity. The starch-statolith hypothesis was also supported by these results.

Interestingly, some other sgr mutants, including sgr2, sgr3, and zig/sgr4, have been shown to still have an endodermis, although the sedimentation of amyloplasts in those cells is abnormal (Morita et al., 2002; Yano et al., 2003). These mutants show a gravitropism defect in the shoot exclusively (inflorescence stem and hypocotyls) and are normal in root gravitropism or phototropism. Surprisingly, responsible genes of these mutants are closely related to the biogenesis or the integrity of the vacuoles (Kato et al., 2002; Yano et al., 2003). Moreover, it is revealed that amyloplasts in the wild-type...
endodermis are almost completely enwrapped by a thin cytoplasmic layer and vacuolar membrane, whereas amyloplasts are left in the peripheral region of the cytoplasm of the endodermal cells in the mutants (Morita et al., 2002). When expressed under the control of the SCARECROW promoter (ProSCR), which drives expression in the endodermis, the corresponding wild-type genes restored gravitropism and amyloplast sedimentation in the mutant (Morita et al., 2002; Yano et al., 2003). These findings suggest that the early step of the gravitropism occurring in the endodermis is impaired in the mutants. They still retain, however, the ability for the latter steps, signal formation in the gravity perceptive cell, intracellular and intercellular signal transduction, and an asymmetric cell elongation between the upper and lower sides of the responding organs for tissue bending. These findings also suggest that vacuoles in the endodermis may be involved in the early step of gravitropism to some extent.

In the gravity-perceptive cell of the root (columella cell), several detailed analyses of the subcellular dynamics have been performed after gravistimulation by reorientation (Legue et al., 1997; Fasano et al., 2001; Yoder et al., 2001), and it has been shown that amyloplasts actually sediment to the orientation of gravity (Yoder et al., 2001). In the shoot, however, only a few attempts have been made at visualizing the gravity-perceptive cell at a subcellular level (Johannes et al., 2001), and to our knowledge, no studies have tried to analyze the movement dynamics of the vacuolar membrane in the endodermal cell. Here, we describe amyloplast movement in the living endodermal cell of Arabidopsis inflorescence stems using a vertical stage microscope. Interestingly, they do not sediment statically but show saltatory movement. We also visualize the movement dynamics of the vacuolar membrane in the endodermal cells. Confocal laser scanning microscopy revealed where amyloplasts pass through. Treatment with a microfilament-disrupting drug severely affected the movement of amyloplasts before gravistimulation by reorientation, although the gravity-perceptive ability of the inflorescence stem segment still remained. In zig-1/sgr4-1 mutants, the movement of amyloplasts was severely affected, and their gravitropism was almost completely defective as shown previously. We will discuss the possible role of the gravity-perceptive system in the dynamics of amyloplasts and the close interaction with the vacuolar membrane. This study opens the possibility of a new concept for gravity perception in the inflorescence stem.

RESULTS

Methods for the Observation of Organelle Dynamics and Subcellular Events in a Living Endodermal Cell

A transgenic plant containing green fluorescent protein (GFP)-marked amyloplasts in its endodermis (pspt3-6) was generated by transforming into wild type an expression construct carrying a GFP sequence fused to a plastid-targeted transit peptide, under the control of ProSCR (Figures 1A and 1B). Amyloplasts were still detectable using autofluorescence even in the non-transgenic plant (wild type); however, a clearer image of amyloplasts was obtained in the transgenic line (Figure 1E). The images are comparable to those made from a fixed sample section (Figures 1C and 1D). ProSCR and GFP were further applied for the visualization of the vacuolar membrane. GFP and γ-TIP (tonoplast intrinsic protein γ), a marker of the lytic vacuole (Maurel
et al., 1993), were fused and expressed in wild-type plants under the control of ProSCR. In the transgenic plant (psgt4-1), the vacuolar membrane is visualized within the endodermis specifically with the band-path filter set (Figure 1F; NIBA; Olympus, Tokyo, Japan). As far as we examined, no significant leak of the marker molecules to other layers was detected in the inflorescence stem, and both pspt3-6 and psgt4-1 lines showed a normal gravitropic response (data not shown).

Dynamics of Amyloplasts in Endodermal Cells of the Inflorescence Stem before and after Gravistimulation by Reorientation

First, we wanted to examine the dynamics of amyloplasts using time-lapse imaging. Sequential images were acquired at 10-s intervals for 5 to 20 min. Amyloplasts did not sediment statically but moved within the endodermal cell dynamically, as

![Figure 2](image-url)
shown by a time-lapse movie (see Supplemental Movie 2 online, x100 speed) and the tracing of movement (Figure 2A, right). They moved not only along the peripheral area, but also through the central region of the endodermal cell very frequently. These dynamic movements were observed continuously even after 20 min (up to 40 min maximum) after preparation of samples.

After gravistimulation by reorientation, the amyloplasts changed their localization pattern drastically and some of them reached the lower end of the cell (Figure 2B, 3 min). They did not stay at the basal end but continued to move around (see Supplemental Movie 3 online). All of the movements of amyloplasts were not directly toward the gravity orientation during this short time period. Some amyloplasts appeared to actually move along a newly applied gravity orientation and others not, as shown by the trace of each amyloplast, after gravistimulation by reorientation (Figure 2B, bottom).

Gravity-Oriented Movement of Amyloplasts Occurs during the First Three Minutes after Reorientation

Attempts were made to analyze the movements of amyloplasts quantitatively and statistically. Tracks of some amyloplasts were traced continuously at 10-s intervals using time-lapse images (Figures 2A, right, and 2B, bottom). To extract gravity-orientated movement, each movement at every 10-s interval was traced continuously at 10-s intervals using time-lapse movies (Figures 2A, right, and 2B, bottom). To extract gravity-orientated movement, each movement at every 10-s interval was traced continuously at 10-s intervals using time-lapse images (Figures 2A, right, and 2B, bottom). To extract gravity-orientated movement, each movement at every 10-s interval was traced continuously at 10-s intervals using time-lapse images (Figures 2A, right, and 2B, bottom). To extract gravity-orientated movement, each movement at every 10-s interval was traced continuously at 10-s intervals using time-lapse images (Figures 2A, right, and 2B, bottom). To extract gravity-orientated movement, each movement at every 10-s interval was traced continuously at 10-s intervals using time-lapse images (Figures 2A, right, and 2B, bottom). To extract gravity-orientated movement, each movement at every 10-s interval was traced continuously at 10-s intervals using time-lapse images (Figures 2A, right, and 2B, bottom). To extract gravity-orientated movement, each movement at every 10-s interval was traced continuously at 10-s intervals using time-lapse images (Figures 2A, right, and 2B, bottom). To extract gravity-orientated movement, each movement at every 10-s interval was traced continuously at 10-s intervals using time-lapse images (Figures 2A, right, and 2B, bottom). To extract gravity-orientated movement, each movement at every 10-s interval was traced continuously at 10-s intervals using time-lapse images (Figures 2A, right, and 2B, bottom). To extract gravity-orientated movement, each movement at every 10-s interval was traced continuously at 10-s intervals using time-lapse images (Figures 2A, right, and 2B, bottom). To extract gravity-orientated movement, each movement at every 10-s interval was traced continuously at 10-s intervals using time-lapse images (Figures 2A, right, and 2B, bottom). To extract gravity-orientated movement, each movement at every 10-s interval was traced continuously at 10-s intervals using time-lapse images (Figures 2A, right, and 2B, bottom). To extract gravity-orientated movement, each movement at every 10-s interval was traced continuously at 10-s intervals using time-lapse images (Figures 2A, right, and 2B, bottom). To extract gravity-orientated movement, each movement at every 10-s interval was traced continuously at 10-s intervals using time-lapse images (Figures 2A, right, and 2B, bottom). To extract gravity-orientated movement, each movement at every 10-s interval was traced continuously at 10-s intervals using time-lapse images (Figures 2A, right, and 2B, bottom). To extract gravity-orientated movement, each movement at every 10-s interval was traced continuously at 10-s intervals using time-lapse images (Figures 2A, right, and 2B, bottom). To extract gravity-orientated movement, each movement at every 10-s interval was traced continuously at 10-s intervals using time-lapse images (Figures 2A, right, and 2B, bottom). To extract gravity-orientated movement, each movement at every 10-s interval was traced continuously at 10-s intervals using time-lapse images (Figures 2A, right, and 2B, bottom). To extract gravity-orientated movement, each movement at every 10-s interval was traced continuously at 10-s intervals using time-lapse images (Figures 2A, right, and 2B, bottom).

| Table 1. Averaged Speed of Amyloplasts (μm/min) Extracted to Δx and Δy Components |
|---------------------------------|---------------------------------|---------------------------------|
| Wild Type                       | +LatB                           | zgt-1/sgr4-1                    |
| Δx                              | Δy                              | Δx                              | Δy                              | n    |
| Before                          |                                  | Before                          |                                  |      |
| –0.1 ± 5.9                     | –0.2 ± 6.7                      | 0.12 ± 2.1                      | 0 ± 2.2                         | 825  |
| After applying gravistimuli     |                                  |                                  |                                  |      |
| 0 to 1 min                      | 0.03 ± 5.5                      | 1.05° ± 5.4                     | 17                              | 117  |
| 1 to 2 min                      | –0.1 ± 4.6                      | 1.42° ± 6.1                     | –87                             | 87   |
| 2 to 3 min                      | 1.05° ± 3.6                     | 1.11° ± 5.4                     | –0.44 ± 2.9                     | 76   |
| 3 to 4 min                      | 1.21° ± 5.6                     | –0.4 ± 4.5                      | 0.41° ± 3.1                     | 90   |
| 4 to 5 min                      | –1.2 ± 4.8                      | –0.1 ± 5.3                      | –0.31 ± 1.9                     | 73   |

The averaged speed was calculated from the same data set that was used in the histograms shown in Figures 2D, 4E, and 5D (cell number, n = 3, total tracks used in the calculation was represented in column n). The axis was defined as shown in Figure 2C. For the direction of movements along Δx and Δy, positive values mean right or down, and negative values indicate left or up, respectively. ±, standard deviations. Asterisks show the time period of the mean values of Δx or Δy statistically significantly increased compared with the mean value of Δx or Δy before gravistimulation by reorientation (Welch’s t test, one-tailed, P < 0.05). LatB, latrunculin B.

Amyloplasts Are in the Transvacuolar Strand, Both before and after Gravistimulation by Reorientation

It has been revealed using electron microscopy that the amyloplasts in the endodermal cells are surrounded almost completely by a thin layer of cytoplasm and the vacular membrane (Morita et al., 2002). The narrow areas of cytoplasm enclosed by the vacular membrane and continuous to the peripheral cytoplasm have been referred to as “transvacuolar strands.” It was expected that amyloplasts move through these transvacuolar strands before or after gravistimulation by reorientation. Vacular membranes in the endodermal cell are visualized well by expressing the GFP-γ-TIP fusion molecule driven under ProSCR. As shown in Figure 1F, the vacular membrane in the endodermal cell emitted fluorescence in the psgt4-1 line, but the strand-like structure was not observed so
clearly using epifluorescence microscopy. However, using a high-speed confocal scanner unit (CSU-10; Yokogawa, Tokyo, Japan), a complex network of the transvacuolar strands and some dilated areas, where amyloplasts are expected to localize, were observed both before (Figure 3A) and after (Figure 3B) gravistimulation by reorientation. From time-lapse confocal laser scanning images, vacuolar membranes were also shown to be quite dynamic even before gravistimulation by reorientation (see Supplemental Movies 4 and 5 online). A silhouette of the vacuolar membrane often showed the same size, shape, and number of amyloplasts in the endodermal cell. These images strongly suggest that amyloplasts move within the transvacuolar strands both before and after gravistimulation by reorientation.

To confirm that amyloplasts translocate through transvacuolar strands, we visualized both amyloplasts autofluorescing and the vacuolar membrane (by GFP-γTIP expression) in the same cell of the psgt4-1 line with a very short time difference by changing the filter set. Figure 3C shows time-lapse images of the vacuolar membrane and amyloplasts observed alternately at 10-s intervals. Most of the GFP-γTIP signals were detected very close to the autofluorescence of amyloplasts. These images clearly indicate that amyloplasts move within the cell wrapped by the vacuolar membrane.

The Saltatory Movement of Amyloplasts Is Microfilament Dependent

To address the molecular basis of the movement of amyloplasts and to examine their association with gravitropism, the effect of the disruption of actin cytoskeleton was analyzed.

After a 10-min treatment with the antimicrofilament drugs cytochalasin B and latrunculin B, the saltatory movements of amyloplasts were severely attenuated (data not shown). As reported previously (Yamamoto and Kiss, 2002), latrunculin B–treated inflorescence stems are able to respond to gravity. These findings were confirmed in this study after 14 h of treatment with latrunculin B (Figures 4A and 4B). It was also shown that the network of actin filaments was completely disrupted, as detected by Pro35SGFP talin (Figure 4B, right). The motility of amyloplasts was considerably reduced, and almost all of them were located at the basal region of the endodermal cell (Figure 4C; see Supplemental Movie 6 online). After gravistimulation by reorientation, however, some populations of amyloplasts actually sedimented, whereas others remained static (Figure 4D; see Supplemental Movie 7 online). Statistical analysis showed that the distribution of ∆y components became steeper, indicating that the movements of amyloplasts were inhibited before gravistimulation by reorientation (Figures 2B and 4E). Positive ∆y components showing large absolute values were detected (asterisks, >3 μm/10 s), and this population was 0.8, 2.5, 1.0, 0, and 0% in 0 to 1 min, 1 to 2 min, 2 to 3 min, 3 to 4 min, and 4 to 5 min, respectively (before reorientation, 0%). The average values of ∆y components were comparable to those of the wild type (Table 1: 1.72 μm/min, 0.94 μm/min, 0.76 μm/min, and 0.86 μm/min in 0 to 1 min, 1 to 2 min, 2 to 3 min, and 3 to 4 min, respectively). The mean values of ∆y during this period are statistically significantly increased compared with the mean value of ∆y before gravistimulation by reorientation (Welch’s t test, one-tailed, P < 0.05). The mean values of ∆x in the time periods of 0 to 1 min and 3 to 4 min are also statistically significantly increased.
Figure 4. The Disruption of the Actin Cytoskeleton and Saltatory Movement of Amyloplasts and Retention of Gravitropism after Treatment with Latrunculin B.

(A) and (B) Gravitropism assay of stem segment and F-actin filament in an endodermal cell visualized by GFP-talin without (A) or with (B) latrunculin B treatment. Left top, 0 min after gravistimulation by reorientation; left bottom, 150 min after gravistimulation by reorientation; right, actin microfilament visualized by Pro35S-driven GFP-talin expression. Bars = 5 μm.

(C) Left, time-lapse images of amyloplasts in the endodermal cell of latrunculin B–treated stem segment at 1-min intervals before gravistimulation by reorientation. Right, trace of amyloplasts. Each color represents the trace of an individual amyloplast. Bars = 5 μm.
compared with the mean value of $\Delta x$ before gravistimulation by reorientation (Welch’s $t$ test, one-tailed, $P < 0.05$). Amyloplasts were still covered almost entirely by the vacuolar membrane (Figure 4F). The effect of a microtubule-disrupting drug (amiprophosmethyl) on the movements of amyloplasts was further examined. The saltatory movements of amyloplasts were not affected by the treatment with amiprophosmethyl, and the stem showed normal gravitropic response (data not shown).

**Dynamic Movement of Amyloplasts Is Severely Restricted in sgr2, sgr3, and zig/sgr4 Mutants Both before and after Gravistimuli**

Among sgr mutants, sgr2, sgr3, and zig/sgr4 were selected to observe the dynamics of amyloplasts in the endodermis of the inflorescence stem because these genes are likely to function in the biogenesis or the integrity of the vacuolar membrane (Kato et al., 2002; Yano et al., 2003). Electron microscopy studies have revealed that amyloplasts in the endodermis of the inflorescence stem of these sgr mutants did not sediment normally, nor were they wrapped by a vacuolar membrane (Morita et al., 2002; Yano et al., 2003). The movement of amyloplasts in these mutants is severely restricted in the endodermal cells of the inflorescence stem. Figures 5A and 5B show the behavior of amyloplasts in the endodermal cell of zig-1/sgr4-1 as a representative result from these mutants (see also Supplemental Movies 8 and 9 online). The amyloplasts in the endodermal cell of the mutants never moved through the central region of the cell, although they occasionally moved a very short distance in the peripheral area of the cell. To examine the defects of the vacuolar membrane in the endodermal cell, the GFP-$\gamma$-TIP fusion construct was expressed in zig-1/sgr4-1 and the stems were subjected to microscopic observation. As a result, transvacuolar strand-like structures were not observed in the endodermal cell. In addition, many abnormal vesicle-like structures accumulated in these cells (Figure 5C), which occasionally showed very dynamic movement (see Supplemental Movie 10 online). Statistical analysis of the movement of amyloplasts in zig-1/sgr4-1 showed that before gravistimulation by reorientation, the distribution of $\Delta y$ and $\Delta x$ components became steeper, indicating that the movements of amyloplasts were inhibited (Figures 2D and 5D). After gravistimulation by reorientation, a small shift of $\Delta y$ components to the positive occurred, but asymmetry of the distribution to the axis of $x = 0$ was not significant. Neither mean values of $\Delta x$ nor $\Delta y$ in all time periods are statistically significantly increased compared with the mean values before gravistimulation by reorientation (Welch’s $t$ test, one-tailed, $P < 0.05$). Average values of the $\Delta y$ components were also calculated (Table 1), and from this the value in zig-1/sgr4-1 was found to be noticeably reduced (0.46 $\mu$m/min, 0.19 $\mu$m/min, and $-0.4 \mu$m/min in 0 to 1 min, 1 to 2 min, and 2 to 3 min, respectively). The occurrence of the $\Delta y$ positive components with large absolute values was not detected in zig-1/sgr4-1.

**DISCUSSION**

**Subcellular Dynamics of the Endodermis in the Arabidopsis Inflorescence Stem**

In this study, the subcellular dynamics of the gravity-perceptive cell in the inflorescence stem of Arabidopsis were observed using fine time and image resolution, with several intriguing findings being noted. First, saltatory movement of amyloplasts was shown both before and after gravistimulation by reorientation in the living graviperceptive cell. Quantitative analysis of a large data set obtained from fine time-lapse imaging revealed that the amyloplasts actually moved toward the direction of gravity 0 to 3 min after gravistimulation by reorientation in the wild-type cell (Figure 2, Table 1). Saltatory movements of amyloplasts in the living graviperceptive cell have also been reported in the dandelion (Taraxacum officinale) flower stalks (Clifford and Barclay, 1980), mung bean (Vigna radiate) hypocotyls (Heathcote, 1981), and the maize (Zea mays) coleoptile (Sack and Leopold, 1985). Rapid movements of amyloplasts toward the gravity were detected (>3 $\mu$m/10 s, $\sim$20 $\mu$m/min) as previously reported in the living graviperceptive cell; however, amyloplasts undergoing such movements constituted only a fraction of the overall population. Averaging the data sets of the amyloplast movement shows that gravity-oriented movement is attenuated by the upward movement component, although the movement can still be detected at $\sim$1 $\mu$m/min. It is plausible that both kinds of gravity-oriented movement are actually occurring 0 to 3 min after gravistimulation by reorientation (Table 1, Figure 2D). These findings are consistent with the presentation time previously reported (Fukaki et al., 1996a).

The vacuolar membrane was visualized and revealed that the translocation path of the amyloplasts occurs in transvacuolar strands, thin cytoplasmic areas that penetrate through the central vacuole. Sack and Leopold (1985) suggested that amyloplasts may move through transvacuolar strands. This study showed using confocal laser scanning microscopy that amyloplasts in the peripheral area of the cytoplasm were almost entirely covered with a vacuolar membrane. The vacuolar membrane itself seems to be dynamic, but it is continuously bent and deformed by

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**Figure 4.** (continued).

(D) Top, time-lapse images of amyloplasts in the endodermal cell of latrunculin B–treated stem segment at 10-s intervals after gravistimulation by reorientation. Bottom, traces of amyloplasts. Each color represents the trace of an individual amyloplast. Bars = 5 $\mu$m.

(E) Histograms showing the frequency of the speed of amyloplasts tracks, extracted to $\Delta x$ and $\Delta y$ components in a wild-type stem segment treated with latrunculin B for 14 h. Total number subjected to the analysis before gravistimulation by reorientation (tracks $n = 404$, cells $n = 3$) and after gravistimulation by reorientation (tracks $n = 520$, cells $n = 4$).

(F) A confocal laser scanning microscopic image of the vacuolar membrane in the endodermal cell of a latrunculin B–treated stem segment. Bar = 5 $\mu$m.
the amyloplasts as well. As shown in Figures 1F and 3C, the amyloplasts are always closely attached and enwrapped by the vacuolar membrane. This interaction between vacuolar membrane and amyloplasts may represent one of the most striking differences between the subcellular organization of the root and shoot graviperceptive cells. In root collumera cells, amyloplasts are not enwrapped by the vacuolar membrane and are thought to move within the cytoplasm with little or no interaction with the vacuole. These findings are consistent with the observation that sgr2, sgr3, and zig/sgr4 mutants still have normal gravitropism in the root, in spite of their shoot gravitropism defects.

Amyloplast Movement for Gravity Perception Is Likely to Be Actin Cytoskeleton Independent

The actin cytoskeleton network has been characterized well in many plant species, with the thick actin cable often observed within the transvacuolar strands (Traas et al., 1987; McCurdy et al., 1988; Tominaga et al., 2000). This study also clearly showed the actin cytoskeleton in the transvacuolar strands and its involvement with the movement of amyloplasts. Findings from the actin disruption experiment suggest that the saltatory movement of amyloplasts through transvacuolar strands is dependent upon the actin cytoskeleton.
The actin cytoskeleton has been proposed as a major player in plant gravitropism (Yoder et al., 2001; Blancaflor, 2002; Hou et al., 2003). Recent studies, however, showed that an intact actin cytoskeleton is not required for gravity perception in inflorescence stems and roots (Yamamoto and Kiss, 2002; Hou et al., 2003) and likely acts to downregulate gravitropism by continuously resetting the gravitropic-signaling system (Hou et al., 2004). The former finding was confirmed in this study, and in addition, the movement of amyloplasts was shown to correlate well with gravity-perceptive ability. In the actin disrupted cell, saltatory movements decline considerably both before and after gravistimulation by reorientation. After gravistimulation by reorientation, translocation of amyloplasts toward the gravity occurs, as shown by both time-lapse imaging and quantitative analysis (Figure 4, Table 1; see Supplemental Movies 6 and 7 online). A few of the amyloplasts can show dramatic and significant movement, even though most of the plastids do not move after reorientation. Although we cannot conclude from these observations that the movement of the amyloplasts was enhanced after actin disruption, as suggested by Yamamoto and Kiss (2002), we can say that the mass translocation of amyloplasts in the stem is comparable to the wild type. These results suggest that the movement of amyloplasts for graviperception is not actin dependent and that the saltatory movement of amyloplasts is actin dependent and not essential for graviperception.

**What Is the Importance of Movement of Amyloplasts for Graviperception?**

In this study, amyloplast dynamics were compared using three types of inflorescence stem: drug-untreated wild type, latrunculin B–treated wild type, and zig-1/sgr4-1 mutant. The former two are graviperceptive, and the latter is a nongraviperceptive stem. The results are summarized in Figure 6. These findings suggest two kinds of movement showing good correlation between gravity-perceptive ability. The first kind is the mass translocation of amyloplasts toward gravity. The average speed of the gravity-oriented movement of amyloplasts in the latrunculin B–treated sample is comparable to that of the wild-type sample, whereas that of zig-1/sgr4-1 is greatly reduced (Table 1). These findings may indicate that mass translocation of amyloplasts toward the lower end of the cell induces gravity signal transduction and therefore is important for graviperception. The second kind of movement is displayed by a population of amyloplasts that sediment at high velocity (>3 μm/s; Figures 2 and 4, asterisks). That population is very small, a few percent, but they may retain a large amount of kinetic energy, taking into account that the kinetic energy is proportional to the square of the velocity, and this large amount of kinetic energy may be enough to overcome the threshold to elicit the signal for a downstream response. This kind of movement may correspond with the sedimentation of amyloplast “faster and to a greater magnitude,” as discussed by MacCleery and Kiss (1999) or “the leading amyloplasts” (Sack et al., 1984). The mean values of Δx direction in some time periods (in wild type, 2 to 3 and 3 to 4 min, and in the latrunculin B–treated samples 0 to 1 min and 3 to 4 min are reorientation) are also statistically significantly increased compared with the mean values before reorientation. The possible explanation for this observation is that amyloplasts tend to move toward the previous upper side of the cell because of the larger area than the previous bottom side of the cell.

**What Is the Role of the Complex Vacuolar Configuration—For Making Paths for Amyloplasts or Other Functions?**

The complex feature of the vacuole in the endodermal cell is presumed to have a function in gravitropism of the inflorescence stem. Because this close interaction of vacuolar membrane with amyloplasts was also reported in cress hypocotyls (Volkmann et al., 1993), this feature may be common in the shoot (in hypocotyls and inflorescence stems). Two possibilities of the function of vacuolar membrane in graviperception are proposed. First, it is proposed that the transvacuolar strands may facilitate smooth translocation of amyloplasts. In the wild type, amyloplasts are translocated within transvacuolar strands; however, in the zig/sgr4 mutant, amyloplasts cannot pass through the transvacuolar strands. Second proposal is that the vacuolar membrane itself may receive the signal directly from the gravity-oriented movement of amyloplasts, for example, through the change of tension in the membrane as a result of gliding amyloplasts. However, using this hypothesis, it is difficult to explain two factors. How is the noisy signal eliminated from continuously moving amyloplasts, and how do we know whether the signal coming from the vacuolar membrane within the cell is upward and/or downward. Because, at present, the vacuolar membrane itself seems not to have any particular polarity. Further careful study is required to address these questions.
METHODS

Sources and Constructions of GFP Marker Lines

The GFP (S65T) with plastid transit peptide (Pt-GFP) of RBCS1A (Chiu et al., 1996; Niwa et al., 1999) was provided by Yasuo Niwa (Niwa et al., 1999; University of Shizuoka, Japan). A BamHI-EcoRI fragment containing Pt-GFP and Nos terminator was then inserted into the BamHI and EcoRI sites of pBI121del_pSCR (Morita et al., 2002) downstream of ProSCR. The GFP (S65T) fused with γ-tubulin intrinsic protein (GFP-γ-TIP) was provided by Hiroshi Abe (RIKEN, Wako, Saitama, Japan). The XbaI-EcoRI fragment containing GFP-γ-TIP and Nos terminator was inserted into the XbaI and EcoRI sites of pBI121del_pSCR (Morita et al., 2002) downstream of ProSCR. The Pro_560-GFP-talin fusion construct was provided by Nam-Hai Chua (Kost et al., 1998; The Rockefeller University, New York, NY).

The constructs were transformed into Agrobacterium tumefaciens strain MP90 by electroporation using the Gene pulser (Bio-Rad, Hercules, CA) and then introduced into wild-type Columbia plants using the floral dipping method (Clough and Bent, 1998). The T1 plants were selected with resistance to kanamycin. The construction of the transgene in these plants was tested using PCR. Segregation of the transgene in the T2 generation was confirmed. The representative lines were selected from T3 homozygous lines by checking the GFP fluorescence in the endodermis of the inflorescence stems. The transgenes were transferred to sgr mutants by transfection (GFP-γ-TIP in zig-1/sgr4-1) or crossing (Pt-GFP in sgr2, agr3, and zig-1/sgr4-1).

Plant Materials and Growth Conditions

In this study, pspt3-6 and psgt4-1 were used as a ProSCR-Pt-GFP and a ProSCR-GFP-γ-TIP expressing line, respectively. Seeds were imbibed, sawed on MS medium (1 × MS salt mixture, 1% [w/v] sucrose, 0.01% [w/v] myoinositol, and 0.5% [w/v] gellan gum, pH 5.8) vernalized in the dark at 4°C for 1 to 2 d, and then grown at 23°C under continuous light. Seedlings were then transplanted to soil to grow inflorescence stems 7 to 14 d after germination. Only the first inflorescence stem was used in the experiments.

Imaging System for Vertical Stage Microscope

An epifluorescence microscope (BX-50; Olympus) equipped with rotary stage was mounted on its back so that the rotary stage was oriented vertically (Legue et al., 1997). Each endodermal cell in the inflorescence stem was observed while it was vertical, and its gravitropic response was elicited by rotating the stage through 90° until the cell layer was horizontal. The endodermal cell was observed before and after gravistimulation. GFP fluorescence or autofluorescence of amyloplasts was acquired with a cooled CCD camera (COOL SNAP cf; Photometrics, Nippon Roper, Tokyo, Japan) and processed using IPLab software (Scanalytics, Fairfax, VA). For confocal laser scanning microscopic observation, a confocal scanner unit (CSU-10; Yokogawa) was mounted to the vertical stage microscope. Images were captured by a CCD camera (ORCA-ER; Hamamatsu Photonics, Hamamatsu, Japan) and processed with Meta-morph (Universal Imaging, Downingtown, PA) and IPLab.

Sample Preparation for the Vertical Stage Microscope

Pieces of cover glass (~170 μm thick; Matsunami, Tokyo, Japan) were cut into ~5-mm-wide strips. These strips were stuck to a glass slide as a spacer using a waterproof adhesive agent (ARON ALPHA; TOAGOSEI, Tokyo, Japan). Double-sided adhesive tape was prepared using two sheets of an adhesive tape stuck together by ARON ALPHA. The double-sided adhesive tape was cut very thinly and stuck onto the spacer. Another piece of the double-sided adhesive tape was stuck inside the spacer, and an inflorescence stem segment (~1 cm in length, the region of 1 to 2 cm from the top of the inflorescence stem) was excised and stuck to the tape. The segment was cut longitudinally using a razor blade held with forceps. Immediately after cutting, a small amount of GM liquid medium (1 × MS salt mixture and 1% sucrose) with 0.1% agar was added. The space inside the well was covered with a cover slip and sealed with double-sided adhesive tape (see Supplemental Figure 1 online).

Electron Microscopy

The samples of inflorescence stem were embedded in Spurr’s resin (Nissin EM, Tokyo, Japan) following the methods of Morita et al. (2002). Ultrathin sections were cut (70 to 90 nm thick), stained with uranyl acetate and lead nitrate, and examined with a transmission electron microscope (H-7100; Hitachi, Tokyo, Japan).

Statistical Analysis of the Amyloplast Movement

The movement of amyloplasts was subjected to quantitative analysis using time-lapse images. Amyloplasts that could be traced for a substantial period (approximately more than five frames) were selected. Each coordinate of the amyloplast in the image frame was recorded, and Δx and Δy was calculated at 10-s intervals. The axis of the coordinate was defined as shown in Figure 2. Pseudomovement was caused by the directional and negligible stage drifting was corrected using the coordinate of an immobile marker (for example, the corner of the cell) in the first and last frames. Traces of the tracks were drawn using a representative example of each condition (before or after gravistimulation by reorientation in the wild type, zig-1/sgr4, and latrunculin B–treated stem segment). The frequency of Δx and Δy at each 10 s interval was presented in a histogram.

Drug Treatment

Stock solutions of latrunculin B (Calbiochem, La Jolla, CA; 20 mM) and amiprophosmethyl (Nihon Bayer Agrochem, Tokyo, Japan; 200 mM) were prepared in dimethyl sulfoxide. A stock solution of Cytochalasin B (Wako, Tokyo, Japan; 20 mM) was prepared in distilled water. First, drugs were added directly into the GM liquid medium. Latrunculin B and cytochalasin B effectively inhibited the saltatory movement of amyloplasts in the endodermal cell in 2 and 0.2 μM, respectively. The drug incorporation experiment was performed according to the methods of Fukaki et al. (1996a), with some modification. Distal 4-cm stem segments of primary inflorescence stems with total lengths between 4 and 12 cm were used for gravitropic response assays. The distal 4-cm parts were cut from the primary inflorescence stems with a razor blade, and their basal ends were inserted into a 1.5-mL microfuge tube positioned vertically on a tube rack. Several stem segments were soaked together in one tube so that they stood in a vertical position. They were preincubated for 14 to 16 h in white light at 23°C in GM liquid medium without sucrose. After incubation, basal 5-mm pieces of these stem segments were put into GM gellan gum blocks that were fixed onto one face of plastic plates (Fukaki et al., 1996a). To maintain a high humidity in the plates, a wet paper towel was placed inside the plate. After the stem segments were incubated in a vertical orientation for 1 h under white light at 23°C in the plates, gravistimulation was initiated by rotating the plates through 90° in darkness at 23°C. Images were taken at the start point and after 150 min using the COOL SNAP cf camera.
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