PLASTID PROTEOME ASSEMBLY WITHOUT TOC159: PHOTOSYNTHETIC PROTEIN IMPORT AND ACCUMULATION OF N-ACETYLATED PLASTID PRECURSOR PROTEINS


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Import of nuclear-encoded precursor proteins from the cytosol is an essential step in chloroplast biogenesis that is mediated by protein translocon complexes at the inner and outer envelope membrane (TOC). Toc159 is thought to be the main receptor for photosynthetic proteins, but lacking a large-scale systems approach, this hypothesis has only been tested for a handful of photosynthetic and nonphotosynthetic proteins. To assess Toc159 precursor specificity, we quantitatively analyzed the accumulation of plastid proteins in two mutant lines deficient in this receptor. Parallel genome-wide transcript profiling allowed us to discern the consequences of impaired protein import from systemic transcriptional responses that contribute to the loss of photosynthetic capacity. On this basis, we defined putative Toc159-independent and Toc159-dependent precursor proteins. Many photosynthetic proteins accumulate in Toc159-deficient plastids, and, surprisingly, several distinct metabolic pathways are negatively affected by Toc159 depletion. Lack of Toc159 furthermore affects several proteins that accumulate as unprocessed N-acetylated precursor proteins outside of plastids. Together, our data show an unexpected client protein promiscuity of Toc159 that requires a far more differentiated view of Toc159 receptor function and regulation of plastid protein import, in which cytosolic Met removal followed by N-terminal acetylation of precursors emerges as an additional regulatory step.

INTRODUCTION

Assembly of the chloroplast proteome requires transport complexes that recognize plastid precursor proteins in the cytosol and mediate their translocation into the chloroplast. Most of the nuclear-encoded chloroplast proteins contain a cleavable N-terminal transit peptide and are imported by the TOC/TIC (for translocon at the outer/inner membrane of the chloroplast) import machinery (Agne and Kessler, 2009; Balsera et al., 2009). In Arabidopsis thaliana, recognition and selection of chloroplast-imported proteins are mediated by GTP binding proteins that belong to two small families: Toc34/33 and Toc159/132/120/90. Members of the Toc159 family share a GTP binding domain (G-domain) and a membrane anchoring domain (M-domain). They are distinguished by differently sized acidic domains (A-domain), with Toc90 entirely lacking the A-domain. The differently sized A-domains in Toc159, Toc132, and Toc120 are located at the N-terminal side of the G- and M-domains and largely account for the molecular mass differences between the different members of the Toc159 family (Agne and Kessler, 2009). The receptors are required for photosynthetic growth because their loss-of-function produces plants with a pale green or albino phenotype (Jarvis et al., 1998; Bauer et al., 2000). Reverse genetic studies and precursor binding assays suggested two different classes of receptors with specialized functions:
one class comprising Toc159/90 and the other class Toc132/120 (Hiltbrunner et al., 2004; Ivanova et al., 2004; Kubis et al., 2004; Smith et al., 2004; Infanger et al., 2011). The loss of either Toc132, Toc120, or Toc90 did not result in a visible phenotype, but Toc132 Toc120 double mutants appeared pale green (Kubis et al., 2004) or were even embryo lethal (Ivanova et al., 2004), suggesting functional overlap between Toc132 and Toc120. Neither full-length Toc132 nor Toc120 could complement ppi2 (Ivanova et al., 2004; Kubis et al., 2004). Based on these data, it was proposed that Toc132 and Toc120 are specific for the import of nonphotosynthetic proteins (Kubis et al., 2004). In contrast with Toc132 and Toc120, the loss of Toc90 function in the Toc120 background resulted in a more severe phenotype, suggesting functional overlap between Toc159 and Toc90 (Hiltbrunner et al., 2004). Indeed, partial complementation of the ppi2 phenotype was achieved by the overexpression of Toc90 (Infanger et al., 2011).

Although it is currently unclear how the different Toc receptors recognize their substrates, it is conceivable that amino acids in the N-terminal transit peptide of plastid precursor proteins are involved in the recognition and in establishing different import specificities. Canonical chloroplast targeting requires transit peptides that are typically 20 to 100 amino acids long and enriched in hydroxylated amino acids but low in acidic amino acids (Bruce, 2001). After import, most transit peptides are cleaved by a stromal processing peptidase (SPP; Richter and Lamppa, 1998, 1999). Processing of imported precursors is essential, and the lack of SPP results in embryo lethality in Arabidopsis (Trösch and Jarvis, 2011). Because the SPP cleavage site sequence is not highly conserved, it was proposed that recognition requires physicochemical properties of the transit peptide rather than a particular amino acid sequence (Emanuelsson et al., 2000; Zhang and Glaser, 2002; Rudhe et al., 2004). Following transit peptide removal, the newly generated protein N terminus may be acetylated (Ferro et al., 2003; Kleffmann et al., 2007; Zybalov et al., 2008). N-acetylation is catalyzed by N-terminal acetyltransferases in the cytosol and in the plastid (Sherman et al., 1985; Meinnel et al., 2006; Goetze et al., 2009). It is therefore possible that a chloroplast-targeted protein may be acetylated as a precursor in the cytosol and after import and processing as the mature protein. N-terminal acetylation is thus a useful modification to map the different N termini of proteins in different compartments. Interestingly, the lack of a cytoplasmic N-acetyltransferase affects plastid biogenesis suggesting a functional crosstalk between cytosolic N-acetylation and plastid function (Pesaresi et al., 2003).

The assembly and maintenance of the chloroplast proteome are highly regulated processes and involve not only the organelar import machinery but also multiple anterograde and retrograde signaling events that affect the transcription of nuclear- and plastid-encoded genes (Mochizuki et al., 2001, 2008; Strand et al., 2003; Larkin et al., 2003; Koussetzky et al., 2007; Moulin et al., 2008; Kakizaki et al., 2009). Similar transcript patterns observed under different environmental conditions or genetic modifications identified key regulators that affect entire sets of plastid proteins and biochemical networks (Richly et al., 2003; Biehl et al., 2005). In order to understand chloroplast proteome assembly, it is therefore necessary to investigate the complexity and quantitative nature of the proteome together with the regulation of the corresponding genes.

Here, we report an integrated approach to investigate the qualitative and quantitative contribution of the Toc159 receptor to the assembly of the plastid proteome. We analyzed the proteome and the transcriptome of Toc159-deficient plants to discern the consequences of impaired protein import from systemic transcriptional responses resulting from the loss of photosynthetic capacity. The analysis of N-terminal protein acetylation showed that some chloroplast-targeted precursor proteins accumulate outside of plastids in Toc159-deficient plants. Interestingly, these precursors were not exclusively photosynthetic proteins but have diverse metabolic functions. Several of these proteins were acetylated at position 2, indicating that they entered the common modification pathway for cytosolic proteins. Our results show that Toc159 has broad specificity, which is different from the proposed role as a receptor for import of abundant photosynthetic proteins. Together, our combined proteome and transcriptome analysis of mutants lacking Toc159 provides new insights into nuclear–organelle interactions during chloroplast proteome assembly.

RESULTS

Quantitative Analysis of Protein Accumulation in Wild-Type and Toc159-Deficient Leaves

To investigate the contribution of Toc159 to the assembly of the plastid proteome, we measured the quantitative protein accumulation in two Arabidopsis mutant lines deficient in Toc159 in comparison to the wild type and the wild type grown on Suc (wtS) (Figures 1A to 1D). The Toc159 null mutant ppi2 (Bauer et al., 2000) can grow beyond the cotyledon stage only on medium supplemented with Suc (Figure 1A). Elevated levels of soluble sugars interfere with developmental processes and affect gene expression (Bilsing et al., 2005; Gibson, 2005). To bypass the requirement of exogenous sugar supply, we generated a Toc159 cosuppression line (Toc159cs) that develops silencing of the Toc159 gene only during later stages of development, resulting in normal appearing seedlings and older plants with chlorophyll-deficient white rosette leaves (Figure 1B). Toc159cs lines were originally constructed to overexpress the M- and G-domains of Toc159 in a wild-type background (see Supplemental Figure 1 online) but developed the cosuppression phenotype shown in Figure 1A. Several independent Toc159cs lines were selected containing only one single transgene insertion. Thermal asymmetric interlaced PCR was used to identify lines in which the transgene was inserted in a noncoding region (see Supplemental Figure 1D online). Immunoblotting analyses with Toc159 antibodies confirmed the complete loss of Toc159 in ppi2 and in white leaves of Toc159cs (Figure 1E).

Using tandem mass spectrometry (MS/MS), we analyzed the leaf proteome composition of wild-type, wtS, ppi2, and Toc159cs plants in three biological replicates each. In total, 4582 different proteins were identified (wild type, 2760; wtS, 2882; ppi2, 3150; Toc159cs, 3119) (Figure 1F; see Supplemental Data Set 1 online). Quantitative information was obtained for all identified proteins using normalized spectral counting (nSpC) (Baerenfaller et al., 2008). Since our experiment was designed to
analyze the consequences of Toc159 depletion for the assembly of the quantitative plastid proteome, we grouped the proteomics data of the wild type (wild type and wtS) and the Toc159-deficient plants (ppi2 and Toc159cs) and compared the protein identification and accumulation within and between these two groups. In total, 2347 proteins were detected in the wild type and wtS and 2496 in ppi2 and Toc159cs. The Spearman rank correlation coefficients obtained from the mean nSpC within these two groups are 0.85 for the wild type and wtS and 0.69 for ppi2 and Toc159cs. Although a large number of proteins were identified in all four plant types (1755 proteins), their quantitative accumulation was considerably different between the wild-type and the Toc159-deficient plants with Spearman rank correlation coefficients between 0.43 and 0.47.

Plastid Proteome Assembly in the Absence of Toc159

To investigate the changes in the abundance of plastid proteins, we extracted these from the leaf data sets using a high-confidence chloroplast proteome reference table comprising 1155 proteins (Baginsky and Gruissem, 2009; see Supplemental Data Set 2 online). We identified in total 926 plastid proteins in our data sets. In Toc159-deficient plants (the ppi2 mutant and Toc159cs), we identified 767 plastid proteins and 569 were identified in all four profiled plant types (Figure 2A). The quantitative accumulation of plastid proteins was very similar between wild-type and wtS plastids and between ppi2 and Toc159cs, with Spearman rank correlation coefficients of 0.945 and 0.813, respectively, and statistical tests support this conclusion. Out of 737 plastid proteins identified with more than 10 spectra, only 27 were significantly different between wild type and wtS and only 22 between Toc159cs and the ppi2 (t test, P value < 0.05). Only two proteins are present in both sets of significantly changed proteins: ferritin (AT5G01600) and β-amylose (AT4G17090); thus, we have no indication for a systematic influence of Suc on quantitative plastid protein accumulation. These results indicate that effects of exogenous Suc supply on plastid proteome assembly are minor under our experimental conditions and that both growth
conditions, with and without Suc, can be used for a quantitative comparison of the plastid proteomes in wild-type and Toc159-depleted plastids. Furthermore, statistical testing (t test, P value < 0.05) revealed that the abundances of the different components of the protein import machinery were not significantly affected by Suc, suggesting that similar constraints exist for the assembly of the plastid proteome in ppi2 and Toc159cs (see Supplemental Figure 2 online).

Next, we used the quantitative data for 737 plastid proteins of which 680 are nuclear encoded that were identified with at least 10 spectra across all samples to identify groups of plastid proteins whose accumulation is significantly affected by the

Figure 2. Quantitative Accumulation of Plastid Proteins Identified in Leaves. (A) Venn diagram of the plastid proteins identified in leaves by MS/MS. Plastid localization is based on a chloroplast proteome reference table with 1155 proteins. wt, wild type. (B) Differential accumulation of nuclear-encoded plastid proteins in wild-type and Toc159-deficient leaves. Numbers have been rounded to one decimal. (C) Metabolic annotation of the nuclear-encoded genes destined to the plastid based on their distribution in MapMan bins. Categories indicated with an asterisk are significantly overrepresented, showing P values < 1e-05 in Fisher’s exact test. CHO, carbohydrate; OPP, oxidative pentose phosphate.
absence of Toc159. For statistical analysis, we omitted proteins identified with <10 spectra because low-abundance proteins are not accurately quantified (Zybailov et al., 2009). We performed a t test and defined proteins that accumulated to higher levels in wild-type or in Toc159-deficient plants as those proteins that had a P value < 0.05 both in the comparisons of the wild type against Toc159cs and wtS against ppi2, and no significant difference in the comparisons of the wild type against wtS and Toc159cs against ppi2 to exclude those proteins from the analysis whose accumulation is affected by Suc. In addition, we required a minimum fold change above 1.5 to exclude spurious small changes that are statistically significant from the interpretation. To these proteins we added those that had been identified in at least two replicates in each wild-type condition and not at all in both Toc159-deficient conditions and vice versa. Altogether, 184 plastid proteins accumulated to higher levels in wild-type and 24 in Toc159-deficient plants, of which 155 and 24, respectively, are nuclear encoded (Figure 2B; see Supplemental Data Set 3 online). The vast majority of the identified plastid proteins are not significantly affected by Toc159 depletion (74%). Two groups of plastid proteins (“no significant change” and “up in Toc159-deficient”) include proteins whose accumulation is not significantly affected by the absence of Toc159. Functional annotation of these putative Toc159-independent proteins and Fisher’s exact test revealed a significant enrichment of the categories photosynthesis (“no significant change”) and tricarboxylic acid (TCA) cycle TCA/org. transformation (“up in Toc159-deficient”) (Figure 2C). By contrast, the third group of proteins, “up in wild type,” comprises 155 nuclear-encoded plastid proteins that accumulate to higher levels in wild-type plastids and thus comprises putative Toc159-dependent substrates (Figures 2B and 2C). Functional annotation showed that these 155 proteins belong to various functional categories (Figure 2C) with a significant enrichment in photosynthesis.

Proteins for Photosynthetic Functions Are Downregulated at the Transcriptional Level in ppi2

To distinguish proteins whose accumulation is directly affected by a loss of Toc159 function from those that are downregulated at the transcriptional level, we profiled the transcriptome of wild-type and ppi2 plants using plants grown on Suc to account for a possible effect of Suc on gene expression. After RNA hybridization to AGRONOMICS1 genome tiling arrays (Rehrauer et al., 2010), signals above background in at least one of the samples were detected for 24,639 of the ∼30,000 probe sets corresponding to 24,378 genes. Of these genes, 1998 (8.2%) were significantly upregulated in the wild type and 2268 (9.3%) in ppi2 (Figure 3A; see Supplemental Data Set 4 online). Our transcriptomics data confirmed the trend that photosynthetic proteins are expressed at lower levels in the import mutant as previously reported in published microarray, SAGE and RT-PCR data (Kubis et al., 2003; Kakizaki et al., 2009; Lee et al., 2009b). Notably, 84% of the downregulated genes reported in the SAGE data set were also found downregulated under our experimental conditions (see Supplemental Data Set 4 online). Similarly, we could confirm the reported upregulation of several heat shock–related proteins (Kakizaki et al., 2009). Of the 1046 nuclear-encoded genes coding for plastid proteins for which we had array data, 381 (36.4%) had significantly reduced RNA levels in ppi2, with a significant overrepresentation of genes encoding proteins for photosynthetic functions (Figure 3B; see Supplemental Data Set 4 online). Thus, the decreased accumulation of photosynthetic proteins in ppi2 is most likely a composite effect of reduced protein import and downregulation of genes for photosynthetic proteins, possibly as a systemic response to the loss of photosynthetic capacity.

We observed that 104 (expected in a random distribution are 59, P value in Fisher’s exact test = 2.2e-07) of the set of 155 nuclear-encoded plastid proteins with decreased protein accumulation in Toc159-deficient plants for which we had transcript levels were also downregulated at the transcriptional level in ppi2 (69.8%; Figures 3B and 3C; see Supplemental Data Set 3 online). For the 24 proteins that accumulated to higher levels in Toc159-deficient plants (Figure 3C), 75% showed no significant change in RNA expression level, 20.8% were upregulated in ppi2 compared with the wild type, and only one was downregulated. From the 529 proteins (Figure 2B; i.e., the 74% plastid proteins from the category “no significant change”), more than 80% were either unchanged or downregulated at the transcriptional level in ppi2, thus indicating that they are efficiently imported into plastids despite the absence of Toc159. These Toc159-independent proteins also include proteins involved in photosynthesis (Figure 2C). In conclusion, the comparison between protein and transcript accumulation therefore confirms that Toc159 is not exclusively involved in the import of photosynthetic proteins and that many photosynthetic proteins enter the plastids in the absence of Toc159 (Figure 2C).

Peptide Detection Incidence Indicates Plastid Precursor Protein Accumulation outside of Plastids in Toc159-Deficient Plants

The accumulation of plastid proteins in Toc159-deficient leaves does not necessarily indicate their correct import into the chloroplast. We therefore analyzed the peptide detection incidence along the amino acid sequence of each protein to distinguish between mature plastid proteins and unprocessed precursors. For each identified protein, the most N-terminal detected peptide was considered and placed into the indicated amino acid bin corresponding to its location in the protein (Figure 4). After import, transit peptides of plastid proteins are usually cleaved off and degraded. Therefore, peptides from the transit peptide region should not be detectable from mature plastid proteins. The bin distribution of peptide detection reveals a detection gap at the N-terminal region of plastid proteins, thus suggesting that the transit peptides of most of the plastid proteins were cleaved (Figure 4A). The distribution was similar between the wild type, wtS, ppi2, and Toc159cs, suggesting that almost all precursor proteins were imported into plastids and processed in the absence of Toc159. As a control, we analyzed the peptide detection incidence for nonplastid proteins (Figure 4B). In this case, peptide detections from the N-terminal region were more prevalent, indicating that the low peptide detection in the N terminus of plastid proteins is most likely a consequence of N-terminal processing.
Figure 3. Transcriptional Response in ppi2 Compared with wtS.

(A) Expression data for 24,639 genes of ppi2 compared with wtS grown on Suc (all genes) and for the subset of 1046 nuclear-encoded plastid proteins. The percentage of downregulated genes in ppi2 highly increases when only nuclear-encoded genes destined to the plastid are taken into account.

(B) Metabolic annotation of the nuclear-encoded genes destined to the plastid based on their distribution in MapMan bins. Categories indicated with an asterisk are significantly overrepresented, showing P values < 1e-05 in Fisher’s exact test.

(C) Expression data for the genes for which the proteins were found significantly upregulated in wild type (wt)/wtS, unchanged, or significantly upregulated in toc159cs/wtS in the proteomics data. CHO, carbohydrate; OPP, oxidative pentose phosphate.
cleaved and degraded. Their detection in ppi2 and Toc159cs therefore provides a first hint that precursor proteins accumulate in these two genotypes (Table 1). To gain further insights into processing of chloroplast-targeted proteins in the wild type and in the mutants, we systematically analyzed N-terminal protein acetylation.

Plastid Precursor Proteins Accumulate outside of Plastids in Toc159-Deficient Plants Where They Are Modified by N-Terminal Met Excision and N-Terminal Acetylation

The detection of peptides within the N-terminal transit peptide sequence of several plastid proteins suggests their accumulation as unprocessed precursors in Toc159-deficient plants. However, the exact size of the transit peptide is difficult to predict and therefore could be erroneous. For further information about precursor processing, we therefore mapped the in vivo N termini of proteins by analyzing N-terminal protein acetylation (N-acetylation) (see Supplemental Data Set 6 online). Since N-acetylation occurs in the cytosol and in plastids, it allows us to distinguish between plastid precursor and mature proteins (Figure 5A). Cytosolic proteins are predominantly acetylated at position 2, which corresponds to the detection of N-acetylated peptides in the 1-10 amino acid bin (Figure 5B). This distribution was similar among the wild type, wtS, ppi2, and Toc159cs, indicating that cytosolic N-terminal Met excision (NME) and subsequent N-acetylation is functional in all plant lines (Meinnel et al., 2006).

By contrast, differences were found in the distribution of N-acetylation sites in plastid proteins. In the wild type, peaks of N-acetylated peptides were detected between amino acid positions 30 and 80, with a minor peak in the 1-10 amino acid bin (Figure 5A). N-acetylation between positions 30 and 80 corresponds to the position of the mature N terminus after removal of transit sequences and is therefore characteristic for imported and processed proteins. The minor peak at positions 1 to 10 comprises four nuclear-encoded proteins, a dynamin-like protein (AT5G42080), a protein of unknown function (AT5G01750), a malate dehydrogenase (AT5G11670), the Fe-superoxide dismutase 1 (AT4G25100), and three plastid-encoded proteins (see Supplemental Data Set 7 online). The nuclear-encoded proteins do not have predicted transit sequences, which is supported by our data.

For ppi2 and Toc159cs plastid proteins, two N-acetylation peaks were visible: a major peak at positions 1 to 10 and a second, minor peak at positions 30 to 80 (Figure 5A). As discussed above, the peak at positions 30 to 80 is indicative for precursor processing after import in ppi2 and Toc159cs plastids. The major peak at positions 1 to 10 in Toc159-deficient plants comprises three plastid-encoded proteins and 16 nuclear-encoded proteins (see Supplemental Data Set 7 online). Ten of these nuclear-encoded plastid proteins have a canonical transit sequence of a predicted length between 29 and 75 amino acids (Table 1). The accumulated proteins are not restricted to abundant photosynthetic proteins but also include minor constituents of the chloroplast proteome, such as two 29-kD ribonucleoprotein, the chaperonin 20, and a thiazole-requiring enzyme. Notably, the sequence context around the identified N-acetylation sites is consistent with established
requirements for the cytosolic NME/acetylation pathway (i.e., small, uncharged residues, such as Ala following the initiator Met) (Figures 5C and 5D) (Sherman et al., 1985; Meinnel et al., 2006). The detection of unprocessed plastid precursor proteins in the import-deficient plants could result either from a cleavage and processing defect in the mutant plastids or from precursor protein accumulation outside of plastids, most likely in the cytosol. The data presented in Figure 6 show that the amino acid context of the identified N-acetylation sites of processed plastid proteins was similar in the wild type and mutants. Although it is possible that a defect in the plastid protease network could potentially affect precursor processing in plastids (Zybailov et al., 2009), our data show that the accumulation of proteins involved in the posttranslational protein homeostasis network is not negatively affected by a defect in Toc159 (see Supplemental Data Set 8 online). Most of the N-acetylated mature plastid proteins that we identified here start with Ala or Val, and Ile and Val are overrepresented at the position three amino acids N-terminal to the N-acetylation site in the precursor protein. This is true for the mature proteins in all genotypes tested and confirms previous data (Zybailov et al., 2008). Together, these data argue against a cleavage and processing defect in ppi2 and Toc159cs lines. We therefore tested whether the detection of unprocessed precursors results from their accumulation outside of plastids using isolated plastids.

### Proteome Analysis with Isolated Plastids Confirms the Accumulation of Precursor Proteins outside Plastids in Toc159-Deficient Mutants

To confirm the accumulation of N-acetylated plastid precursors outside of plastids in Toc159-deficient plants, we isolated plastids from wild-type and ppi2 plants because unprocessed cytosolic precursors should be removed during plastid isolation. We analyzed quantitative protein accumulation in isolated wild-type and ppi2 plastids using MS/MS in three biological replicates and identified 3231 unique proteins (see Supplemental Figure 3 and Supplemental Data Set 9 online). We focused our analysis on 856 proteins that were present in the chloroplast proteome reference table (Baginsky and Gruissem, 2009). Of these proteins, 708 were identified in the wild type, 708 in ppi2, and 589 in both

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**Table 1. Unprocessed Precursor Proteins Accumulate outside Plastids in Toc159-Deficient Plants**

<table>
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<tr>
<th>Accession</th>
<th>Description</th>
<th>N-Acetylation Position (Leaves)</th>
<th>Minimal Peptide Position in Leaves</th>
<th>Toc159 (Wild Type)</th>
<th>Toc159 Wild Type</th>
<th>AtProteome</th>
<th>TP</th>
<th>Length</th>
<th>Metabolic Function</th>
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<td>130</td>
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<td>68</td>
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<td>–</td>
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<td>54</td>
<td>86</td>
<td>Secondary met.</td>
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Cytosolic location of plastid proteins was assessed by detection of N-acetylation within the predicted transit peptide (TP). Provided is also the minimal position of detected peptides in wild-type leaves and in AtProteome (Baerenfaller et al., 2008). Data “Wild Type” encompass data from wild type and wr5, and “toc159” data from ppi2 and Toc159cs, respectively. CHO, carbohydrate; NF, not found; met., metabolism; org., organic acid; -, no transit peptide predicted.
genotypes (see Supplemental Figures 3A and 3B online). The high number of proteins common to both plastid types provides additional evidence that most plastid proteins accumulate in both wild-type and ppi2 plastids despite the lack of Toc159. The N-terminal peptide detection incidence of isolated wild-type and ppi2 plastids was very similar, indicating that the transit sequence of most plastid proteins was cleaved after import (see Supplemental Figure 3C online). Also, the amino acid context around the identified N-acetylation sites of wild-type and ppi2 plastid proteins was in agreement with the previous observation for all leaf data sets, confirming that processing of imported plastid proteins is functional in the wild type and in ppi2 plastids (Figure 6; see Supplemental Data Set 9 and Supplemental Figure 3D online).

Analysis of acetylated N termini supports the import and correct processing of plastid precursor proteins in isolated wild-type and in Toc159-deficient leaves. The major peak in bin 1-10 in leaves was absent in isolated plastids, suggesting that cytosolic precursor proteins were removed during plastid isolation (see Supplemental Data Set 10 online). Since peptide detection in data-dependent acquisition may result in false-negative results, we searched for the peptide masses of N-terminal peptides from precursor proteins in the leaf data set and in isolated plastids. The absence of detectable peptide masses in MS spectra provides a strong argument against its presence in the sample because the mass spectrometric measurement is extremely sensitive. Most peptides of a complex sample will give rise to a data point (i.e., peak) in the spectrum, even though only a very small fraction of these are identified in data-dependent acquisition experiments (Beck et al., 2011). While the precursor peptide masses were clearly identified in leaf extracts of Toc159-deficient plants, no masses matching the precursor peptide were identified in isolated plastids (exemplified for At3g12780 and At1g61520 in Figure 8 and Supplemental Figure 4 online). If nuclear-encoded plastid precursor proteins were present in plastids, the isolation procedure should have enriched these compared with the leaf extract and, thus, they should be more readily detectable. Based on these data, we therefore conclude that the unprocessed precursor proteins in ppi2 and Toc159cs accumulate outside of plastids.
plastids. The typical processing pattern of cytosolic proteins that is also observed for the detected precursor proteins (i.e., NME and N-terminal acetylation) (Figure 5) furthermore provides a strong argument for their cytosolic localization.

Identification of Toc159-Dependent and Toc159-Independent Plastid Proteins

Using the quantitative protein and transcript data presented above, we defined substrates that depend on Toc159 for import into the chloroplast and searched for features that distinguish such Toc159-dependent and Toc159-independent plastid proteins. We defined as Toc159-dependent those proteins that accumulated to significantly higher levels in the wild type and wtS compared with ppi2 and Toc159cs leaves as described above (see Supplemental Data Set 3 online). To exclude those proteins whose decreased abundance in ppi2 could be attributed to lower transcript levels, we additionally excluded those from the list of Toc159-dependent proteins that were significantly downregulated in ppi2 compared with wtS. As a final criterion, plastid proteins annotated by PPDB and AT_CHLORO to be localized in the outer envelope were removed (Sun et al., 2009; Ferro et al., 2010). Together, 44 proteins fulfilled these requirements and constitute the set of Toc159-dependent proteins, whose accumulation is affected at the protein and not at the transcript level (see Supplemental Data Set 11 online). By contrast, Toc159-independent proteins are those proteins that have increased or equal protein abundance in ppi2 or Toc159cs compared with the wild type and wtS, while their expression levels are similar in ppi2 and wtS. Together, 308 Toc159-independent proteins matched these criteria (see Supplemental Data Sets 1 and 3 online). Most of these were also identified in isolated ppi2 plastids, further supporting their Toc159-independent import (see Supplemental Data Set 9 online).
Among the Toc159-independent proteins was plastoglobulin 35 (PGL35) (At4g04020) that we selected as an additional control for Toc159-independent protein import. The PGL35 protein can be considered representative for this group of proteins because it accumulates to similar levels in wild-type and albino leaves (average nSpC values are as follows: wild type, 1.35; wtS, 1.16; ppi2, 1.22; Toc159cs, 1.46) (see Supplemental Data Set 1 online). We fused PGL35 to green fluorescent protein (PLG35:GFP) (Vidi et al., 2006) and bombarded wild-type and white Toc159cs leaves with the fusion construct attached to gold particles (Figure 9). We observed the accumulation of PLG35:GFP in several wild-type plastids and in smaller plastids of white Toc159cs leaves (Figure 9A). By contrast, the precursor protein of the small subunit of ribulose-1,5-bis-phosphate carboxylase/oxygenase fused to GFP only accumulated in wild-type plastids (Figure 9B). These data confirm our proteomics strategy for the quantification and selection of Toc159-independent proteins.

To extract information about sequence features that might govern import specificity, we aligned the sequences of the 100 first amino acids of the putative Toc159-dependent and Toc159-independent substrates (see Supplemental Figure 5 online). The enriched amino acids comprise mostly Ser residues, and the pattern did not reveal any specific sequence or domain structure that would allow distinguishing proteins in the two data sets. Other attempts to define relevant information in the amino acid sequences by multiple sequence alignment using ClustalW were not successful, and none of the approaches allowed us to distinguish the proteins in the two data sets by sequence information.

**DISCUSSION**

**The Chloroplast Proteome in the Absence of Toc159**

We used quantitative proteomics and transcriptomics to investigate the contribution of the import receptor Toc159 to the assembly of the chloroplast proteome. In addition to the loss-of-function mutant ppi2, we used a Toc159 cosuppression line that allowed us to study the chloroplast proteome composition without exogenous sugar supply (Figure 1). We expected that the natural distribution of source/sink relationships in the Toc159cs plants better reflects the in vivo situation compared with growth of ppi2 on Suc. Interestingly, the qualitative and quantitative accumulation of plastid proteins in leaves was very similar in Toc159cs and ppi2, and also in the wild type and wtS, suggesting that exogenous Suc supply has only a minor effect on the assembly of the plastide proteome.

Based on the above data, ppi2 and white leaves of Toc159cs are equivalent with respect to plastid proteome assembly in the absence of Toc159. Both genotypes accumulate photosynthetic proteins (Figures 2, 4, and 6), and proteins from several other metabolic pathways in the plastids of Toc159-deficient leaves are reduced (Figure 2). Thus, the substrate specificity of Toc159 is not restricted to photosynthetic proteins. Most of the proteins that had a lower abundance in ppi2 and Toc159cs were also downregulated at the gene expression level, suggesting that their decreased accumulation is a result of two different
responses, the transcriptional response as well as the import defect (Figure 3). This is especially true for photosynthetic proteins. It could be argued that import deficiency directly affects gene expression, but such a scenario would require separate signaling circuits that control the expression of individual nuclear genes encoding plastid proteins, which seems unlikely. A more likely explanation is that nuclear-encoded plastid proteins are regulated in modules, which is supported by the proposed master switch model (Richly et al., 2003; Biehl et al., 2005).

Many plastid proteins accumulate in Toc159-deficient plants, some of them to similar or even above wild-type levels. Apparently, the absence of Toc159 is partially compensated by one or more of its homologs: Toc132, Toc120, or Toc90. We could not detect these receptors by MS, either in leaves or in isolated plastids, but transcripts of Toc132 and Toc120 were slightly elevated in ppi2 compared with the wild type (see Supplemental Figure 1 and Supplemental Data Set 4 online). A recent study showed that the overexpression of Toc90 can partially restore the accumulation of Toc159 client proteins in ppi2 (Infanger et al., 2011). However, the low abundance of the Toc90 protein and the low expression level of its transcript in ppi2 make it unlikely that the residual protein import observed here is exclusively based on Toc90 activity (Bauer et al., 2000; Infanger et al., 2011). It is more plausible that Toc132 and Toc120 sustain protein import in Toc159-deficient plants because their expression is induced in ppi2. Elevated receptor abundance was previously shown to restore an albino phenotype to a pale-green phenotype, suggesting that receptor abundance is limiting for efficient protein import (Hiltbrunner et al., 2004; Ivanova et al., 2004; Kubis et al., 2004). Together with these observations, our data call for a model in which the import capacity is controlled not only by substrate specificity but also by the abundance of the different receptors and their ability to associate into functional translocon complexes.

A recent report suggested that the A-domain of the Toc159 receptor family mediates precursor selectivity (Inoue et al.,

Figure 8. Transit Peptide Visualization and Extracted Ion Chromatogram Quantification for Phosphoglycerate Kinase1 Peptides Identified in Leaves and Isolated Plastids.

(A) Visualization of peptide detection along the protein. Several spectra matched to the transit peptide region (indicated as TP; 75 amino acids predicted length) in Toc159-deficient (toc159) leaves. By contrast, we could not detect any spectra matching the predicted TP region in wild-type leaves or in isolated ppi2 or wild-type plastids, although the coverage of Phosphoglycerate Kinase1 (PGK1) detection is similar between the different samples (cf. peptide color scheme and peptide localization between the different samples; the darker the shading, the more spectra matching to the indicated peptide were observed).

(B) Extracted exact mass ion chromatogram for the spectra matching to the transit peptide region from leaves (top panel; marked with an asterisk) and corresponding samples from isolated plastids. In three samples from isolated plastids, an ion matching the exact mass of the PGK1 transit peptide identification (asterisk) was detected at a shifted retention time of ~8 min (bottom panels). MS/MS information on these ions revealed that they are clearly different from the peptide eluting at 48 min (top panel, precursor peptide). The upper peptide represents the peptide matching to the PGK1 transit peptide, the three ions below produced a poor-quality MS/MS spectrum that could not be assigned to any peptide (it was assigned to format dehydrogenase with a score below the threshold). L, leaves; P, plastids.

[See online article for color version of this figure.]
which explains why overexpression of full-length Toc132 or Toc120 failed to complement ppi2 (Ivanova et al., 2004; Kubis et al., 2004), while constructs containing only the G- and M-domains of Toc132 were able to do so (Agne et al., 2009; Richardson et al., 2009; Inoue et al., 2010). Loss of the A-domain resulted in import receptors with less selective precursor protein recognition and import capacities. It seems that the A-domain confers precursor selectivity to a constitutive import process, whose efficiency depends mainly on the abundance of the different Toc receptors. Initial results suggest that the A-domain may be cleaved upon assembly of the Toc complex (Agne et al., 2010). The conclusion that the A-domain mediates import specificity could explain why our data did not identify sequence specificity determinants of Toc159-mediated protein import (see Supplemental Figure 5 online). Although we identified information-rich regions within the first 20 amino acids up to position 55 as reported for Toc159-mediated RbcS import (Smith et al., 2004; Lee et al., 2008; Lee et al., 2009a), we could not identify sequence determinants that distinguish Toc159-dependent from Toc159-independent proteins (see Supplemental Figure 5 online). Additional components in the cytosol, such as 14-3-3 proteins, might be involved in determining specific peptide recognition by different TOC receptors, but further research is necessary to establish the exact role of cytosolic guidance complexes in this process (May and Soll, 2000).

**N-Terminal Met Excision and Acetylation of Plastid Precursor Proteins in the Cytosol**

We found several unprocessed precursor proteins that were N-acetylated at position 2 most likely in the cytosol of Toc159-deficient plants (Figure 5). The detection of the unprocessed precursors raises the question of which specific features allow their accumulation in Toc159-deficient plants. The identified proteins are not simply abundant photosynthetic proteins that may saturate the impaired plastid import machinery in ppi2 and Toc159cs and thus accumulate because of their high abundance. The accumulation of most high-abundance photosynthetic proteins seems to be prevented by regulatory signaling circuits that result in reduced expression of their genes (Figures 2 and 3). Interestingly, some of the unprocessed precursors of nonphotosynthetic proteins were also downregulated at the transcriptional level, but their decrease was not as pronounced as that of photosynthetic proteins (Table 1; see Supplemental Data Set 4 online). This suggests that these nonphotosynthetic proteins are integrated in a signaling circuit that is different from the signaling circuit that controls the expression of photosynthetic proteins. Together, our data support the view that the accumulation of nuclear-encoded plastid precursor proteins triggers an unimported precursor protein response (Kakizaki et al., 2009; Lee et al., 2009b). This signaling response could act through additional unknown signaling pathways that do not depend on the functional state of the plastids.

Our data show that plastid precursor proteins were modified by NME and subsequent N-acetylation outside the plastid. This modification mechanism is used for 60 to 90% of all proteins in eukaryotes (Goetze et al., 2009), and it is conceivable that it also modifies plastid precursor proteins under normal wild-type conditions during their cytosolic synthesis and transition. The sequence context around the identified N-acetylation sites is consistent with described requirements for this type of modification (i.e., small, uncharged residues like Ala follow the start Met) (Sherman et al., 1985; Meinzel et al., 2006). Our observation may therefore indicate a general modification pathway for plastid proteins in the cytosol. N-acetylation serves as a degradation signal for the ubiquitin ligase-proteasome system in yeast, and it was proposed to be involved in protein folding quality control (Hwang et al., 2010). Support for a function of plastid precursor protein N-acetylation in vivo came from the characterization of a mutant for a cytoplasmic N-acetyltransferase. This mutant is affected in chlorophyll accumulation and photosynthetic capacity, suggesting crosstalk between cytosolic N-acetylation and chloroplast function (Pesaresi et al., 2003).

The fact that plastid precursor stability is tightly controlled in the cytosol was recently substantiated by the detection of a

![Figure 9](image-url)
ubiquitin-dependent degradation pathway for nonimported Lhcb4 precursors that is mediated by the heat shock cognate protein 70-4 and ubiquitination by an E3 ligase (Lee et al., 2009b). Inhibition of the proteasome resulted in the enhanced accumulation of ubiquitinated Lhcb4 precursors in ppi2, thus indicating the existence of a degradation system for plastid precursors. Based on these observations, it is conceivable that cytosolic N-acetylation may serve as an additional degradation signal for nonimported plastid proteins to avoid accumulation of unprocessed precursor proteins in the cytosol. A short half-life of nonimported N-acetylated plastid precursor proteins could be the reason why these have not been identified in wild-type plants. Future work is needed to establish the role of N-acetylation in the assembly of the plastid proteome and its contribution to cellular function.

In summary, we used quantitative proteomics in combination with transcriptional profiling to establish a definition of Toc159-dependent and Toc159-independent plastid protein import. Based on our results, we could not identify any amino acid sequence specificity determinants that allow distinguishing Toc159-dependent and Toc159-independent proteins. The numerous photosynthetic proteins that accumulate as correctly processed mature proteins in ppi2 and Toc159cs and the distinct functions affected in Toc159-deficient plastids furthermore indicate an unexpected client protein promiscuity of Toc159 and suggest that the current model for the import of photosynthetic and housekeeping nonphotosynthetic proteins must be revised. The identification of N-acetylated plastid precursor proteins outside the plastid in Toc159-deficient plants could point to a control mechanism for plastid precursor proteins that may prevent precursor accumulation under normal wild-type conditions. Altogether, we present evidence for complex regulation of plastid proteome assembly in import-deficient plastids that encompasses import preference, transcriptional regulation, and potentially differential degradation of plastid precursor proteins by N-acetylation–triggered processes. The quantitative contribution of each individual pathway must be assessed under different conditions, and more information on their dynamic regulation is necessary. This information in combination with increasingly sensitive analyses of precursor protein accumulation via detection of unprocessed N termini, possibly by combined fractional diagonal chromatography (Gevaert et al., 2007), in the future could provide a more comprehensive understanding of chloroplast proteome assembly.

METHODS

Plant Material

All experiments used Arabidopsis thaliana plants of accession Wassilewskija. Plants were grown at 20°C in photoperiods of 8 h light and 16 h darkness with a light intensity of 150 μmol m⁻² s⁻¹. For protein extraction, leaves (leaf 6 from 5 to 10 plants per sample) were harvested after 35 d, 1 h before the end of the photoperiod and frozen in liquid nitrogen. The wild-type and Toc159cs plants were grown on soil, whereas wtS and ppi2 were grown on half-strength Murashige and Skoog medium supplemented with 100 mM Suc. For each genotype, three independent biological replicates were performed (i.e., plants were grown three times under exactly the same conditions), and leaf 6 was harvested as described above. The genotypic characterization of ppi2 and Toc159cs was described by Bauer et al. (2000) and in Supplemental Figure 1 online, respectively.

Generation of Toc159 Cosuppression Plants

A fragment from Toc159 corresponding to the GTPase and membrane domains was amplified by PCR using 5′-CAGGATCCCAATGGCTCTAG-GATCACCCACCCACCCACCCACGGACGAGTTTGTCTATGGAT-3′ (includes BamHI) and 5′-GCTCTAGATTAGTACGCTGTTGTCG-TTTGTC-3′ (includes XbaI) primers, digested with BamHI and XbaI, and inserted into the corresponding NcoI and XbaI sites from the pCH7 binary vector (a gift from C. Falkhauer, University of Lausanne, Switzerland). Plants were transformed using the floral dip method as described (Clough and Bent, 1998). Transformants were selected on soil by spraying phosphinothricin (30 mg/L BASTA; Duchefa), and DNA prepared from leaf material was used for diagnostic PCR. Analysis of segregation was performed by germinating seeds on half-strength Murashige and Skoog medium and 30 μg/mL phosphinothricin. Phosphinothricin-sensitive plants displayed yellow cotyledons and were easily distinguishable from Toc159 cosuppressing plants with white leaves and cotyledons. Lines with a segregation indicative of a single T-DNA insertion and with high incidence of Toc159 cosuppression were selected. T-DNA insertions were mapped in these lines by three rounds of thermal asymmetric interlaced PCR using pCH7_LB_a (5′-CGAACATCGTCT-GAATGCAAAG-3′), pCH7_LB_b (5′-CTACCTGCGTCTGGACGAG-3′), pCH7_LB_c (5′-CGGATACCTAGCTACGTTTGGC-3′), and random primers.

Protein Extraction and Immunoblotting

Protein extracts were prepared from leaves by grinding shock-frozen tissue. Subsequently, extraction buffer was added (40 mM Tris, pH 6.8, 40 mM DTT, 4% SDS, and 2% protease inhibitor cocktail [Roche]). Homogenates were centrifuged twice at 20,000 rcf for 20 min at room temperature. Protein concentration was determined using a BCA protein assay kit (Thermo Scientific) before adding DTT. Twenty micrograms of protein from leaves were subjected to SDS-PAGE on 12% gels. Immunoblotting using α-Toc159, α-PGL35, and α-CAB antibodies was performed as described previously (Ivanova et al., 2004). For each immunoblotting experiment, a second gel was prepared in parallel as loading control, which was stained with Coomassie Brilliant Blue according to standard procedures.

Sample Preparation for Large-Scale Proteomics and MS

Protein extraction was performed as described above, and 600 μg of protein from leaves and 200 μg from isolated chloroplasts were subjected to SDS-PAGE on 12% gels. After electrophoresis, gels were stained with Coomassie Brilliant Blue according to standard procedures and cut into 12 sections for leaves and 20 sections for isolated chloroplasts respectively. Each gel slice was diced into small pieces. In-gel digestion was performed according to Shevchenko et al. (1996). After digestion, dried peptides were resuspended in 3% acetonitrile and 0.2% trifluoroacetic acid and desalted using Sepak cartridges (Waters).

Dried peptides were resuspended in 3% acetonitrile and 0.2% formic acid and analyzed with a linear trap quadrupole Fourier transform–ion cyclotron resonance mass spectrometer (ThermoFischer Scientific) coupled to an Eksigent nano liquid chromatography system (Eksigent Technologies). Peptide mixtures were loaded onto laboratory-made capillary columns (B&G Analytik) of 75-μm inner diameter, 8-cm length, packed with 3 μm, 200 Å Magic C18 AQ beads (Michrom BioResources). Peptides were eluted from the column by an increased acetonitrile gradient from 3% acetonitrile, 0.2% formic acid to 36% acetonitrile, 0.2% formic acid over 35 min, and from 36% acetonitrile, 0.2% formic
acid to 80% acetonitrile, 0.2% formic acid over 5 min, followed by a 10-min wash step with 80% acetonitrile and 0.2% formic acid. Peptide ions were detected in a survey scan from mass-to-charge ratio 300 to 1600 at 100,000 full width half maximum nominal resolution followed by three data-dependent MS/MS scans (isolation width 2 atomic mass units, relative collision energy 35%, dynamic exclusion enabled, repeat count 1, followed by peak exclusion for 2 min).

Data Mining and Protein Quantification

MS/MS spectra were searched either with Sequest and Peptide Prophet (Yates et al., 1995; Keller et al., 2002) using the Trans-Proteomic Pipeline (TPP v2.9) or with Mascot version 2.2.04 (Matrix Science) against the Arabidopsis TAIR9 protein database (downloaded on June 29, 2009) with a concatenated decoy database supplemented with contaminants (87,079 entries). For Sequest, the search parameters were as follows: requirement for tryptic ends, one missed cleavage allowed, mass tolerance = ±5 ppm, variable modification of Met (M, PSI-MOD name: oxidation, ModAccession: MOD: 00412, mono Δ = 57.0214) and static modification of Cys (C, PSI-MOD name: iodoacetamide derivative, ModAccession: MOD: 00397, mono Δ = 57.0214). For Peptide Prophet, the cutoff was set to a minimum probability of 0.9. For Mascot, the search parameters were as follows: requirement for semitryptic ends, one missed cleavage allowed, mass tolerance = ±5 ppm. Besides carbamidomethylation of Cys residues as fixed modification, oxidation of Met, N-terminal protein acetylation, and N-terminal acetylation were included as variable modifications. N-terminal acetylated peptides were accepted with a Mascot ion score higher than 24 and a Mascot expectation value smaller than 0.05. All peptide assignments from Sequest and Mascot except those of contaminants were filtered for ambiguity, and the smaller than 0.05. All peptide assignments from Sequest and Mascot were also excluded from further analysis. Furthermore, spectrum assignments to decoy database peptides were excluded. All the same protein were so identified in Toc159-deficient samples, but in at least two of the three replicates both for ppi2 and Toc159cs (category “Only in Toc159cs/ppi2”), and those that were not at all identified in Toc159-deficient samples, but in at least two of the three replicates both for wild-type and wtS (category “Only in wild type/wtS”). All analyses were done with R (R Development Core Team, 2010).

Microarray Hybridization and Quantification

RNA was extracted from 35-d-old plants. The experiment was performed with three biological replicates, and Affymetrix Arabidopsis AGRONOMICS1 microarrays were used. Labeling of samples, hybridizations, and measurements were performed as described (Hennig et al., 2004; Rehrauer et al., 2010). Signal values were derived using the robust multiarray analysis algorithm implemented in the statistical language R (R Development Core Team, 2010) using probe sets comprising exonic probes based on the TAIR9 genome annotation. For details of probe set definition and low-level data analysis, see Rehrauer et al. (2010). Differentially expressed genes were selected using LIMMA (Smyth, 2004) followed by multiple testing correction according to Storey and Tibshirani (2003). Genes were considered as differentially expressed if the P value < 0.05. This resulted in a list of 15,882 genes. Downstream analysis was performed on a subset of the differentially expressed genes that had a fold change of more than 2. Significance of overlaps of gene sets was calculated with a Fisher’s exact test. The microarray data have been submitted to ArrayExpress with the experiment number E-TABM-951.

Chloroplast Localization, Sequence Alignment, and Functional Annotation

Chloroplast localization annotation was based on a chloroplast protein reference table that comprises 1155 proteins (reviewed in Baginsky and Gruissem, 2009). Plastid localization of these 1155 proteins were validated in different proteomics studies by software tools that integrate targeting prediction with coexpression data and several other criteria that were combined and weighted with a naïve Bayesian classifier. Amino acid sequence alignment and visualization were performed with Weblogo (weblogo.berkeley.edu) (Crooks et al., 2004). In silico localization prediction was done using TargetP (Emanuelsson et al., 2000). Functional annotation was based on MapMan (mapman.mpimp-golm.mpg.de) (Usadel et al., 2009), and the overrepresentation of functional categories was assessed with Fisher’s exact test.

Chloroplast Isolation

Intact chloroplasts from 35-d-old wild-type or ppi2 plants grown on soil or on half-strength Murashige and Skoog medium supplemented with 100 mM Suc medium were isolated according to Fitzpatrick and Keegstra (2001) with the following modifications. Cellulase Onozuka R-10 and macerazyme R-10 (Serva; catalog numbers 16419 and 28302) were diluted to 1 and 0.25% (w/v), respectively. L-ascorbate (0.1% [w/v]) and 0.05% (w/v) protease inhibitor cocktail for plant cell extracts (Sigma-Aldrich;
catalog number P9599) was added to the breakage buffer. Intact wild-
type chloroplasts were harvested at the interface of a Percoll step
gradient (40% [v/v] and 85% [v/v]) and ppi2 chloroplasts on a linear gradient
of 10 to 60%. Different density gradient centrifugation protocols were
used for wild-type and ppi2 plastids because they differ in their density and the
linear gradients resulted in improved purity. To minimize contamination
from other cellular compartments, isolated chloroplasts were subjected to
three successive Percoll gradients, including three washings steps.

Expression of GFP Fusions in Wild-Type and Toc159cs Leaves
Expression in wild-type and Toc159cs leaves of a GFP fusion to the transit
peptide of ribulose-1,5-bis-phosphate carboxylase/oxygenase small
subunit (pSSU-GFP) or of a GFP fusion to an N-terminal fragment of
peptide of ribulose-1,5-bis-phosphate carboxylase/oxygenase small
Toc159cs

Supplemental Data
The following materials are available in the online version of this article.

Supplemental Figure 1. Characterization of Toc159 Cosuppression Lines.

Supplemental Figure 2. Protein and Transcript Abundances of the
Components of the TIC/TOC Import Machinery.

Supplemental Figure 3. Proteome Profiling of Isolated Wild-Type
and ppi2 Chloroplasts.

Supplemental Figure 4. Extracted Ion Chromatogram Quantification
for N-Terminal Peptides in Leaves and Isolated Plastids from the
Light-Harvesting Complex Protein LHCA3 (AT1G61520).

Supplemental Figure 5. Toc159-Independent and Toc159-Dependent
Substrates.

Supplemental Data Set 1. Proteins Identified by MS/MS in Wild-
Type, wtS, Toc159cs, and ppi2 Leaves.

Supplemental Data Set 2. Reference List of 1155 High-Confidence
Plastid Proteins.

Supplemental Data Set 3. Statistical Analysis of Differential Protein
Accumulation in Wild-Type and Toc159-Deficient Plants Compared
with Their Transcriptional Regulation.

Supplemental Data Set 4. Transcriptional Profiling Data for wtS and
ppi2 Plants.

Supplemental Data Set 5. Nuclear-Encoded Proteins with a Minimal
Peptide Position between 1 and 10.

Supplemental Data Set 6. N-Acetylated Peptides Identified by MS/
MS in Wild-Type, wtS, Toc159cs, and ppi2 leaves.

Supplemental Data Set 7. Plastid Proteins Identified by MS/MS in
Wild-Type, wtS, Toc159cs, and ppi2 Leaves N-Acetylated between
Positions 1 and 10.

Supplemental Data Set 8. Stromal Proteins Involved in the Plastid
Posttranslational Protein Homeostasis Network (Zybaliov et al., 2008).

Supplemental Data Set 9. Proteins Identified in Isolated ppi2 and
Wild-Type Plastids.

Supplemental Data Set 10. N-Acetylated Peptides Identified by MS/
MS in Isolated Wild-Type and ppi2 Plastids.

Supplemental Data Set 11. Toc159-Independent and Toc159-
Dependent Proteins.

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AUTHOR CONTRIBUTIONS

S. Bischof, K.B, W.G., and S. Baginsky designed the research. S. Bischof,
research. S. Bischof, K.B., L.H., F.K., and S. Baginsky analyzed data. S.
Bischof, W.G., and S. Baginsky wrote the article.

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Supplemental Data
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