

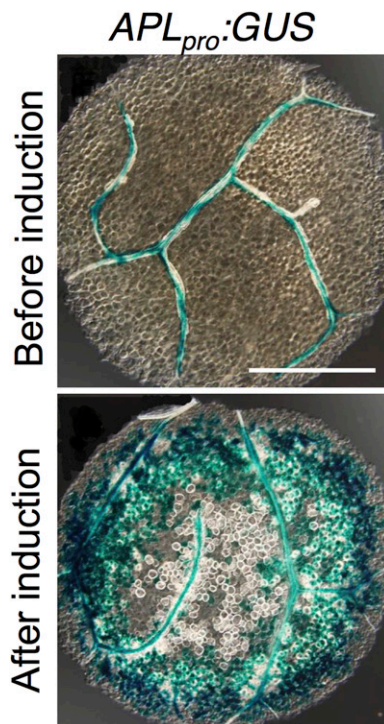
## IN BRIEF

# Thinking Outside the Plant: Exploring Phloem Development Using VISUAL<sup>OPEN</sup>

Investigating how plants grow and develop often requires a bit of creativity. For example, deep within the plant, the vascular cambium, a layer of embryonic, highly cytoplasmic cells, gives rise to xylem and phloem tissue, which must expand throughout the plant's lifetime. Water and nutrients primarily flow through the xylem, whereas photosynthate and signaling molecules travel through the phloem. While anatomical and labeling studies have shed light on the sequential development of vascular tissue, this process is notoriously difficult to track due to the limited accessibility of the plant vasculature. However, under the proper conditions, various cells can be coaxed into producing vascular cells in culture, which greatly facilitates their investigation. Such analyses have revealed many key factors that regulate the differentiation of xylem cells from vascular cambium cells (reviewed in Růžička et al., 2015). However, unlike xylem cells, which undergo marked changes during development, *in vitro*-produced phloem cells are difficult to distinguish from other cell types without the use of phloem-specific markers.

Kondo et al. (2016) developed a powerful system for analyzing phloem differentiation called VISUAL (Vascular Cell Induction Culture System Using Arabidopsis Leaves). In this system, mesophyll cells from *Arabidopsis thaliana* leaf discs or cotyledons are induced to form procambial cells, which then develop into xylem (Kondo et al., 2015) or phloem cells. To unlock the capacity of mesophyll cells to form vascular cells *in vitro*, including sieve element (SE)-like phloem cells and tracheary elements (xylem), the cells are treated with bikinin, an inhibitor of the kinase GSK3, which normally suppresses vascular tissue differentiation. Induction with bikinin triggers the formation of numerous phloem precursor cells and differentiating SEs, as revealed by the increased expression of the phloem markers *APL<sub>pro</sub>:GUS* ( $\beta$ -glucuronidase gene driven by the *ALTERED PHLOEM DEVELOPMENT* promoter; see figure) and

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Expression of the phloem marker *APL<sub>pro</sub>:GUS* in leaf discs before and after induction with bikinin. Bar = 500  $\mu$ m. (Reprinted from Kondo et al. [2016], Supplemental Figures 1F and 1G.)

*SEOR1pro:SEOR1-YFP* (yellow fluorescent protein gene driven by the *SIEVE ELEMENT OCCLUSION-RELATED1* promoter), respectively. DNA inhibitor analysis indicated that cell division is required prior to phloem cell differentiation in VISUAL. *In vitro* and *in vivo* phloem cell differentiation appear to share several crucial steps, such as the formation of SE reticulum, bundles of P-protein filaments, and thick primary cell walls, as well as an enucleation-like event. The induced phloem cells mainly consist of SEs but not companion cells, as shown by microarray analysis. The authors used flow cytometry to isolate phloem SEs expressing *SEOR1pro:SEOR1-YFP*, finding that bikinin-induced mesophyll cells gave rise to 300 times more YFP-positive cells than uninduced mesophyll cells. In addition, phloem-specific genes were

highly expressed in YFP-positive cells, whereas xylem-specific genes were highly expressed in YFP-negative cells.

The authors then took advantage of VISUAL to investigate phloem SE development in an *apl* mutant, which is embryo lethal, since *APL* is a crucial transcription factor. Microarray analysis revealed that *APL* preferentially up-regulates many phloem-specific genes in VISUAL, especially those functioning during later SE development. Next, three different sets of VISUAL transcriptome data, including time-course, *SEOR1* cell-sorting, and *apl* mutant data, were used to construct networks of genes related to SE-like cell differentiation. This analysis identified a transcription factor gene, *NAC020*, that appears to be expressed upstream of *APL*. Indeed, *NAC020* was expressed in *apl*, confirming its function upstream of *APL*. Both overexpression and targeted repression of *NAC020* led to reduced *APL* expression, as well as partially inhibited SE differentiation. These findings point to the importance of *NAC020* in early phloem development, and they hint at the great potential of using VISUAL to explore phloem development outside the plant.

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## REFERENCES

- Kondo, Y., Fujita, T., Sugiyama, M., and Fukuda, H. (2015). A novel system for xylem cell differentiation in *Arabidopsis thaliana*. *Mol. Plant* 8: 612–621.
- Kondo, Y., Nurani, A.M., Saito, C., Ichihashi, Y., Saito, M., Yamazaki, K., Mitsuda, N., Ohme-Takagi, M., and Fukuda, H. (2016). Vascular cell induction culture system using Arabidopsis leaves (VISUAL) reveals the sequential differentiation of sieve element-like cells. *Plant Cell* 28: 1250–1262.
- Růžička, K., Ursache, R., Hejátko, J., and Helariutta, Y. (2015). Xylem development - from the cradle to the grave. *New Phytol.* 207: 519–535.

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