INTRODUCTION

Plant cell walls are amazingly complex amalgams of carbohydrates, proteins, lignin, water, and incrusting substances such as cutin, suberin, and certain inorganic compounds that vary among plant species, cell types, and even neighboring cells. Developmental events and exposure to any of a number of abiotic and biotic stresses further increase this compositional and structural variation. Moreover, the dynamic nature and functions of plant cell walls in terms of growth and development, environmental sensing and signaling, plant defense, intercellular communication, and selective exchange interfaces are reflected in these variations. Much is currently known about the structure and metabolic regulation of the various cell wall components, but relatively little is known about their precise functions and intermolecular interactions.

In this review, I will discuss the accumulated structural and regulatory data and the much more limited functional and intermolecular interaction information on five plant cell wall protein classes. These five protein classes, listed in Table 1, include the extensins, the glycine-rich proteins (GRPs), the proline-rich proteins (PRPs), the solanaceous lectins, and the arabinogalactan proteins (AGPs). These five proteins may be evolutionarily related to one another, most obviously because each of them, with the exception of the GRPs, contains hydroxyproline, and less obviously in the case of the GRPs because this class has nucleotide sequence similarity to the extensins. For completeness, I should mention that these are not the only cell wall proteins that are known. Others exist, such as cysteine-rich thionins, 28- and 70-kD water-regulated proteins, a histidine-tryptophan-rich protein, and many cell wall enzymes such as peroxidases, phosphatases, invertases, α-mannosidases, β-mannosidases, β-1,3-glucanases, β-1,4-glucanases, polygalacturonase, pectin methyltransferases, malate dehydrogenase, arabinosidases, α-galactosidases, β-galactosidases, β-glucuronidases, β-xylodases, proteas, and ascorbic acid oxidase (Varner and Lin, 1989). However, the above five classes generally represent the most abundant, and to date, the most well-studied and widely documented, plant cell wall proteins.

Before describing these five wall protein classes, I should point out that research on these individual proteins has occurred in several plant species, but relatively few examples exist where these cell wall proteins have been studied together in one plant, let alone in one particular plant organ or type of cell. Thus, data from one plant species are often extrapolated to represent the situation in other plant species. Although such extrapolations are usually valid, enough variations are now known that caution should be exercised in making or believing such claims.

EXTENSINS

Structure

Extensins are a family of hydroxyproline-rich glycoproteins (HRGPs) found in the cell walls of higher plants. In dicots, extensins are particularly abundant and are generally characterized by the following: they are rich in hydroxyproline and serine and some combination of the amino acids valine, tyrosine, lysine, and histidine; they usually contain the repeating pentapeptide motif Ser-Hyp, often within the context of other, larger repeating motifs; most of the hydroxyproline residues are glycosylated with one to four arabinosyl residues, and some of the serine residues are glycosylated with a single galactose unit; they are basic proteins with isoelectric points of ~10 due to their high lysine content; they generally assume a polyproline II helical structure in solution; and they have a rodlike appearance when viewed in the electron microscope. These characteristics are exemplified by numerous dicot extensins (Stuart and Varner, 1980; Leach et al., 1982; Mellon and Helgeson, 1982; Smith et al., 1984, 1986; van Holst and Varner, 1984; Cassab et al., 1985; Esquerre-Tugayé et al., 1985; Stafstrom and Staehelin, 1986; Biggs and Fry, 1990; Kawasaki, 1991) and extensin cDNA and genomic clones (Chen and Varner, 1985a, 1985b; Showalter et al., 1985, 1991; Corbin et al., 1987; Showalter and Varner, 1987; Keller and Lamb, 1989; Evans et al., 1990; Gatehouse et al., 1990; Sauer et al., 1990; De Loose et al., 1991; Adams et al., 1992; Zhou et al., 1992).

A recently characterized extensin from sugar beet, a primitive dicot, provides an interesting contrast to the many extensins characterized from advanced herbaceous dicots (Li et al., 1990). This extensin apparently lacks the repeating pentapeptide motif Ser-Hyp, but it does contain a repeating variation...
Table 1. Five Major Classes of Structural Plant Cell Wall Proteins and Some Distinguishing Properties

<table>
<thead>
<tr>
<th>Protein Class</th>
<th>% Protein</th>
<th>% Sugar</th>
<th>Abundant AA(a)</th>
<th>AA Motifs</th>
<th>Selected Prior Reviews</th>
</tr>
</thead>
<tbody>
<tr>
<td>“Extensins” (Chlamydomonas)</td>
<td>25–50</td>
<td>50–75</td>
<td>O,S</td>
<td>SOOOOTOVYK</td>
<td>Cassab and Varner (1988)</td>
</tr>
<tr>
<td>GRPs (dicot)</td>
<td>~100</td>
<td>~100</td>
<td>G</td>
<td>TPKPTOITYTOSOKPO</td>
<td>Showalter and Rumeau (1990)</td>
</tr>
<tr>
<td>GRPs (monocot)</td>
<td>~100</td>
<td>~100</td>
<td>G</td>
<td>PPP and PP</td>
<td>Kieliszewski and Lamport (1988)</td>
</tr>
<tr>
<td>Solanaceous lectins</td>
<td>~55</td>
<td>~45</td>
<td>O,S,G,C</td>
<td>PPVYK and PPVEK</td>
<td>Marcus et al. (1991)</td>
</tr>
</tbody>
</table>

\(a\) O = hydroxyproline; AA = amino acid.

b? = unknown.

of this motif, namely Ser-Hyp\(2[X]\)-Thr-Hyp-Val-Tyr-Lys, where \[X\] represents a Val-His-Glu-Lys-Tyr-Pro “insertion sequence.” Table 2 shows that this sugar beet sequence motif is similar to peptide repeats found in tomato P1, carrot, tobacco, and petunia extensins.

In monocots, somewhat different versions of extensin apparently exist. For example, in the graminaceous monocot maize, both a threonine-hydroxyproline-rich glycoprotein (THRGP) and a histidine-hydroxyproline-rich glycoprotein (HHRGP), which is also rich in alanine, are present (Kieliszewski and Lamport, 1988). The THRGP is particularly well characterized by both protein (Kieliszewski and Lamport, 1987; Kieliszewski et al., 1990) and molecular cloning studies (Stiefel et al., 1988, 1990). Such studies show that the THRGP is distinct from a typical dicot extensin in that it is rich in threonine and proline in addition to hydroxyproline, tyrosine, and serine; it contains two novel amino acid repeat motifs, Thr-Pro-Lys-Pro-Thr-Hyp-Tyr-Thr-Hyp-Lys-Pro-Hyp and Ala-Thr/Thr-Lys-Pro-Pro but only one Ser-Hyp, sequence; the serine residues and approximately half of the hydroxyproline residues are not glycosylated; and it is predicted to exist in a random coil conformation, not in a polyproline II helix. Another, apparently related THRGP is also found in maize, but it has yet to be cloned (Hood et al., 1988). Cloned extensin sequences from two other graminaceous monocots, sorghum and rice, show clear sequence similarity to the known maize THRGP sequence (Raz et al., 1991; Caelles et al., 1992).

Amino acid sequence analysis of a number of chymotryptic peptides of the maize HHRGP has demonstrated the presence of several amino acid sequence repeats including Ala-Hyp\(3\), Ala-Hyp\(4\), and Ser-Hyp\(2-3\) repeats; however, at this point it is still unclear how these repeat motifs are arranged (Kieliszewski et al., 1992b). Recently, another group has cloned an apparent HRGP from maize that is expressed specifically in the pollen. This maize HRGP cDNA clone specifies several Lys-Ser-Ser/Pro-Pro-Ala-Pro-X-Ser-Pro-X repeats, in which X represents some hydrophobic amino acid (A. Broadwater, A. Rubinstein, K. Lowrey and P. Bedinger, personal communication).

Gymnosperms also contain cell wall HRGPs. Examination of salt eluates of Douglas fir cell suspension cultures has revealed an apparent HRGP. One of these HRGPs has sequence characteristics very similar to the PRPs and will be discussed later (Kieliszewski et al., 1992a), whereas the other appears to contain both Ser-Hyp and Ala-Hyp repeat units (Fong et al., 1992), which are typically thought to be characteristic of extensins and AGPs, respectively. Indeed, this latter HRGP is extraordinarily interesting as it tends to blur the distinction between extensins and AGPs and indicates the possible shuffling of diverse repeat domains that may occur in evolution. Bao et al. (1992) have also found that wood from loblolly pine contains a cell wall HRGP with 24% proline and 11% hydroxyproline; amino acid sequences from this interesting protein remain to be elucidated.

Lower plants appear to have yet other versions of extensin or, more generally, HRGPs. For example, the green alga Chlamydomonas contains at least two sets of HRGPs (Adair and Snell, 1990). One set is characteristically distributed in the cell walls of vegetative and gametic cells, whereas the other set is found in the cell walls of zygotic cells. Both sets of HRGPs appear as rodlike molecules in electron micrographs, with the former set containing more bends in the fibrous structure. Further, molecular cloning has shown that the zygotic cells contain
Table 2. Some Intriguing Amino Acid and Nucleotide Sequence Repeat Similarities among the Various Structural Cell Wall Proteinsa

<table>
<thead>
<tr>
<th>Cell Wall Protein</th>
<th>Amino Acid Sequence Repeatb</th>
<th>No. of Repeats</th>
<th>Nucleotide Sequence Repeatc</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Tomato extensis P1</td>
<td>SOO00TOVVK</td>
<td>NDd</td>
<td>NAe</td>
<td>Lamport (1977); Smith et al. (1986)</td>
</tr>
<tr>
<td>Carrot extensis gene</td>
<td>SPPPTPVPVVK</td>
<td>7</td>
<td>TCT CCA CCA CCA CCA ACA CGG GTT TAC TAC AAG</td>
<td>Chen and Varner (1985b)</td>
</tr>
<tr>
<td>Tobacco extensis cDNA</td>
<td>SPPPTPVPVVK</td>
<td>6</td>
<td>TCT CCA CCA CCA CCA ACT CCA GTT TAC AAG</td>
<td>J. Memelink, personal communication</td>
</tr>
<tr>
<td>Petunia extensis cDNA</td>
<td>SPPPTPVPVVK</td>
<td>1</td>
<td>TCC CCA CCA CCA ACC CTA GTT TAC AAG</td>
<td>C. Hironaka, personal communication</td>
</tr>
<tr>
<td>Tomato extensis P1</td>
<td>SOO00VOYPYHTOVTX</td>
<td>ND</td>
<td>NA</td>
<td>Smith et al. (1986)</td>
</tr>
<tr>
<td>Sugar beet extensis</td>
<td>SOO00HYP00TOVTX</td>
<td>ND</td>
<td>NA</td>
<td>Li et al. (1990)</td>
</tr>
<tr>
<td>Tomato extensis P1</td>
<td>SOO00TOVVK</td>
<td>ND</td>
<td>NA</td>
<td>Lamport (1977); Smith et al. (1986)</td>
</tr>
<tr>
<td>Carrot extensis gene</td>
<td>SPPPTPVPVVK</td>
<td>7</td>
<td>TCT CCA CCA CCA CCA ACA CGG GTT TAC TAC AAP</td>
<td>Chen and Varner (1985b)</td>
</tr>
<tr>
<td>Tobacco extensis cDNA</td>
<td>SPPPTPVPVVK</td>
<td>6</td>
<td>TCT CCA CCA CCA CCA ACT CCA GTT TAC AAP</td>
<td>J. Memelink, personal communication</td>
</tr>
<tr>
<td>Petunia extensis cDNA</td>
<td>SPPPTPVPVVK</td>
<td>1</td>
<td>TCC CCA CCA CCA ACC CTA GTT TAC AAP</td>
<td>C. Hironaka, personal communication</td>
</tr>
<tr>
<td>Maize THRG</td>
<td>SPPPTPVPVVK</td>
<td>6</td>
<td>TCC CCA CCA CCA ACC CTA GTT TAC AAP</td>
<td>Stiefel et al. (1988, 1990)</td>
</tr>
<tr>
<td>Maize THRG</td>
<td>SPPPTPVPVVK</td>
<td>5</td>
<td>AGC ACA ACC CGG CCA ACT ACT TAC TAC ACC</td>
<td>Stiefel et al. (1988, 1990)</td>
</tr>
<tr>
<td>Tomato extensis P3</td>
<td>SOO000000000000</td>
<td>ND</td>
<td>NA</td>
<td>Kieliszewski et al. (1990); Stiefel et al. (1988, 1990)</td>
</tr>
<tr>
<td>Tomato extensis cDNA</td>
<td>SPPPPSSCPPPPPPYVK</td>
<td>3</td>
<td>TCC CCA CCA CCA CCA TAC TAC TAC TAC TAC AAP</td>
<td>Showalter et al. (1991)</td>
</tr>
<tr>
<td>Petunia extensis cDNA</td>
<td>SPPPPSSCPPPPPPYVK</td>
<td>3</td>
<td>TCC CCA CCA CCA CCA TAC TAC TAC TAC TAC AAP</td>
<td>C. Hironaka, personal communication</td>
</tr>
<tr>
<td>Bean extensis cDNA</td>
<td>SPPPPSSCPPPPPPYVK</td>
<td>6</td>
<td>TCC CCA CCA CCA CCA TAC TAC TAC TAC TAC AAP</td>
<td>Corbin et al. (1987)</td>
</tr>
<tr>
<td>Bean extensis cDNA</td>
<td>SPPPPSSCPPPPPPYVK</td>
<td>4</td>
<td>TCC CCA CCA CCA CCA TAC TAC TAC TAC TAC AAP</td>
<td>Corbin et al. (1987)</td>
</tr>
<tr>
<td>Maize THRG</td>
<td>TUG00000Y</td>
<td>1</td>
<td>ACC CCC CCC CCC ACC CCA CCA TAC TAC</td>
<td>Kieliszewski et al. (1990); Stiefel et al. (1988, 1990)</td>
</tr>
<tr>
<td>Tomato extensis P1</td>
<td>SOO000000000000</td>
<td>ND</td>
<td>NA</td>
<td>Lamport (1977); Smith et al. (1986)</td>
</tr>
<tr>
<td>Carrot extensis gene</td>
<td>SPPPTPVPVVK</td>
<td>7</td>
<td>TCT CCA CCA CCA CCA ACA CGG GTT TAC TAC AAP</td>
<td>Chen and Varner (1985b)</td>
</tr>
<tr>
<td>Tobacco extensis cDNA</td>
<td>SPPPTPVPVVK</td>
<td>6</td>
<td>TCT CCA CCA CCA CCA ACT CCA GTT TAC AAP</td>
<td>J. Memelink, personal communication</td>
</tr>
<tr>
<td>Petunia extensis cDNA</td>
<td>SPPPTPVPVVK</td>
<td>1</td>
<td>TCC CCA CCA CCA ACC CTA GTT TAC AAP</td>
<td>C. Hironaka, personal communication</td>
</tr>
<tr>
<td>Tomato extensis P2</td>
<td>SOO000000000000</td>
<td>ND</td>
<td>NA</td>
<td>Lamport (1977); Smith et al. (1986)</td>
</tr>
<tr>
<td>Soybean PRP cDNA</td>
<td>PP0YVK</td>
<td>19</td>
<td>CCA ACC --- --- GTW TAC AAG</td>
<td>Averyhart-Fullard et al. (1988)</td>
</tr>
<tr>
<td>Soybean PRP gene</td>
<td>PP0YVK</td>
<td>29</td>
<td>CCA ACC --- --- GTW TAC AAG</td>
<td>Hong et al. (1987)</td>
</tr>
<tr>
<td>Soybean PRP</td>
<td>PP0YVK</td>
<td>ND</td>
<td>NA</td>
<td>Lindstrom and Vodkin (1991)</td>
</tr>
<tr>
<td>Sugar beet extensis</td>
<td>SO000000000000</td>
<td>ND</td>
<td>NA</td>
<td>Li et al. (1990)</td>
</tr>
<tr>
<td>Soybean nodulin PRP cDNA</td>
<td>PP-HEKPP</td>
<td>12</td>
<td>CCA CCT CAT GAG AAP CCA CCA CCA</td>
<td>Datta et al. (1989)</td>
</tr>
<tr>
<td>Soybean PRP cDNA</td>
<td>PPVEA</td>
<td>16</td>
<td>CCA CCT CAT GAG AAP</td>
<td>Fransen et al. (1987)</td>
</tr>
<tr>
<td>Soybean nodulin PRP cDNA</td>
<td>PPHEK</td>
<td>16</td>
<td>CCA CCT CAT GAG AAP</td>
<td>Fransen et al. (1987)</td>
</tr>
<tr>
<td>Tomato extensis P3</td>
<td>SOO000000000000</td>
<td>ND</td>
<td>NA</td>
<td>Lamport (1977); Epstein and Lamport (1984)</td>
</tr>
<tr>
<td>Maize HRGP</td>
<td>A0000HFP500</td>
<td>ND</td>
<td>NA</td>
<td>Kieliszewski et al. (1992a)</td>
</tr>
<tr>
<td>Maize HRGP</td>
<td>A0000</td>
<td>ND</td>
<td>NA</td>
<td>Kieliszewski et al. (1992a)</td>
</tr>
<tr>
<td>Maize HRGP</td>
<td>A0000</td>
<td>ND</td>
<td>NA</td>
<td>Kieliszewski et al. (1992a)</td>
</tr>
<tr>
<td>Maize HRGP</td>
<td>A0000</td>
<td>ND</td>
<td>NA</td>
<td>Kieliszewski et al. (1992a)</td>
</tr>
<tr>
<td>Lolium AGP</td>
<td>SO00APAP</td>
<td>ND</td>
<td>NA</td>
<td>Gleeson et al. (1989)</td>
</tr>
</tbody>
</table>

a Note that eight sets of comparisons are presented, which examine (1) advanced dicot extensis versus a primitive dicot extensis, (2) advanced dicot extensis versus a monocot "extensis," (3) advanced dicot extensis versus a monocot "extensis," (4) advanced dicot extensis versus PRPs, (5) a primitive dicot extensis versus a nodulin PRP, (6) a PRP versus a nodulin PRP, (7) an advanced dicot extensis versus a monocot "extensis," and (8) a monocot "extensis" versus an AGP.

b Note that conserved protein sequences for a given set are underlined, and that gaps (indicated by "-" ) were inserted in some sequences to maximize alignment of the sequences. Also, note that hydroxyproline (O) is indicated only in the case of amino acid studies; amino acid sequences deduced from DNA sequences cannot distinguish proline from hydroxyproline.

c Special nucleotide symbols used include: P = A, G; Q = C, T; R = A, T; S = C, G; W = G, T; 3 = A, C, G, T.

d ND = not determined.

e NA = not applicable (protein study).
an HRGP with (Pro)$_3$ and (Ser-Pro)$_n$ motifs (Woessner and Goodenough, 1989), whereas the vegetative cells contain HRGPs with (Pro)$_3$, Pro-X-Pro, Pro-XX-Pro, and Leu-Pro sequence repeats (Adair and Apt, 1990). Additionally, the plus and minus agglutinins of Chlamydomonas, which bring gametes of opposite mating types together during sexual reproduction, are HRGPs that are similar in nature to the vegetative HRGPs. The sequence Leu-Leu-Hyp-Hyp was found to be present in both sexual agglutinins and in a vegetative HRGP (Adair and Snell, 1990). In Volvox, a novel sulfated, extracellular HRGP, consisting of a globular and a rod-shaped domain, is expressed specifically during embryonic inversion. This glycoprotein was recently cloned, and its rod-shaped domain was found to consist of numerous Ser-Hyp$_3$, repeat units (Ertl et al., 1992).

The evolutionary relationships of the above extensins and the apparent extenin homologs, and hence of the plants that contain them, are now being examined by DNA and protein sequence analysis (Kieliszewski et al., 1990; Showalter and Rumeau, 1990). For example, as shown in Table 2, some intriguing similarities that may indicate interesting evolutionary relationships are seen between (1) the primitive dicot extenin of sugar beet and other dicot extensins; (2) a submotif of one maize extenin repeat, Ser-Hyp-Lys-Pro-Hyp-Thr-Pro-Lys, and the tomato extenin P1 repeat, Ser-Hyp$_2$Thr-Hyp-Val-Tyr-Lys (note that amino acid sequence identity can be achieved with a Lys to Pro conversion and a Val-Tyr deletion); and (3) another maize THRGP repeat, Thr-Hyp-Ser-Hyp$_2$-Tyr, and a motif found in tomato extenin P3, Ser-Hyp-Ser-Hyp$_2$-Tyr (which is a portion of the larger tomato extenin repeat Ser-Hyp$_2$-Ser-Hyp$_2$-Tyr$_2$-Lys). Additional data, however, will have to be obtained before precise evolutionary relationships are elucidated and tenuous links solidified.

Regulated Expression

Various conditions and treatments, as listed in Table 3, generally increase the expression of extenin (reviewed in Showalter and Rumeau, 1990). These studies were performed largely with advanced herbaceous dicots, although maize THRGP expression is regulated during development and by wounding (Stiefel et al., 1988; Ludewid et al., 1990; Fritz et al., 1991; Ruiz-Avila et al., 1991), and Chlamydomonas and Volvox HRGP expression is developmentally regulated (Woessner and Goodenough, 1989; Adair and Snell, 1990; Ertl et al., 1992). Gene activation is probably involved with such changes; however, only in beans (Phaseolus vulgaris) has transcriptional regulation in response to wounding, fungal infection, fungal elicitor, and glutathione been verified by nuclear run-off studies (Lawton and Lamb, 1987; Wingate et al., 1988). Two groups have now reported the existence of nuclear trans-acting factors that interact with specific cis-acting elements of the carrot extenin gene promoter in a wound-specific and ethylene-specific fashion (Holdsworth and Laties, 1989a, 1989b; Granell et al., 1992). Similarly, Wycoff et al. (1991) have carried out promoter deletion experiments with a bean extenin gene promoter fused to the 3-glucuronidase (GUS) reporter gene and demonstrated the existence of cis-acting regions involved in developmental and wound-regulated gene expression.

Some of the most exciting recent work with regard to the extensins, as well as some of the other cell wall proteins, has occurred in the area of cell and tissue localization. Tissue print RNA and protein blots have allowed extenin mRNA and protein to be localized in several plants and tissues. These simple yet elegant tissue print studies have complemented more sophisticated and labor-intensive studies performed using in situ hybridization and immunocytochemical localization. Table 4 shows that taken as a whole, these studies reveal that extenin gene expression and localization can apparently vary from plant to plant and among cell and tissue types, presumably in accord with the different functions of different cell types and tissues. Extensin is commonly associated with phloem tissue and cambium cells, but it can be associated with other tissues as well. In addition, transgenic plants carrying extenin promoters to drive GUS expression have been used to illustrate both tissue-specific and gene-specific expression for extenin (see Table 4).

Although tissue printing is a simple and powerful technique, it is limited by the efficiency of the transfer of protein or mRNA

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**Table 3. Conditions Regulating the Expression of the Five Major Classes of Structural Plant Cell Wall Proteins**

<table>
<thead>
<tr>
<th>Protein Class</th>
<th>Condition(s)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Extensins (dicot)</td>
<td>Wounding, fungal infection, viral infection, fungal elicitors, endogenous elicitors, ethylene, red light, heat shock, gravity, glutathione, cell culturing, development</td>
</tr>
<tr>
<td>&quot;Extensins&quot; (monocot)</td>
<td>Development, wounding</td>
</tr>
<tr>
<td>&quot;Extensins&quot; (Chlamydomonas)</td>
<td>Development</td>
</tr>
<tr>
<td>&quot;Extensins&quot; (Volvox)</td>
<td>Development</td>
</tr>
<tr>
<td>GRPs (dicot)</td>
<td>Development, viral infection, salicylic acid, abscisic acid, drought stress, wounding</td>
</tr>
<tr>
<td>GRPs (monocot)</td>
<td>Development, water stress, abscisic acid, mercuric chloride, wounding</td>
</tr>
<tr>
<td>PRPs</td>
<td>Development</td>
</tr>
<tr>
<td>PRPs (nodulins)</td>
<td>Wounding, endogenous elicitors, fungal elicitor, ethylene, cell culturing, light, red light, development</td>
</tr>
<tr>
<td>Solanaceous lectins</td>
<td>Wounding, viral infection</td>
</tr>
<tr>
<td>AGPs</td>
<td>Development, wounding</td>
</tr>
<tr>
<td>Protein Class</td>
<td>Plant System</td>
</tr>
<tr>
<td>---------------</td>
<td>--------------</td>
</tr>
<tr>
<td>Extensins</td>
<td>Soybean stems and petioles</td>
</tr>
<tr>
<td>Extensins</td>
<td>Bean stems and petioles</td>
</tr>
<tr>
<td>Extensins</td>
<td>Tomato stems and petioles</td>
</tr>
<tr>
<td>Extensins</td>
<td>Petunia stems</td>
</tr>
<tr>
<td>Extensins</td>
<td>Tobacco stems</td>
</tr>
<tr>
<td>Extensins</td>
<td>Carrot roots</td>
</tr>
<tr>
<td>Extensins</td>
<td>Soybean roots</td>
</tr>
<tr>
<td>Extensins</td>
<td>Tomato root</td>
</tr>
<tr>
<td>Extensins</td>
<td>Tobacco with bean extensin-GUS transgene</td>
</tr>
<tr>
<td>Extensins</td>
<td>Rape with rape extensin-GUS transgene</td>
</tr>
<tr>
<td>Extensins</td>
<td>Soybean seeds</td>
</tr>
<tr>
<td>Extensins</td>
<td>Maize (i.e., THRGP)</td>
</tr>
<tr>
<td>GRPs</td>
<td>Soybean stems and petioles</td>
</tr>
<tr>
<td>GRPs</td>
<td>Bean stems</td>
</tr>
<tr>
<td>GRPs</td>
<td>Bean stems</td>
</tr>
<tr>
<td>GRPs</td>
<td>Bean stems</td>
</tr>
<tr>
<td>GRPs</td>
<td>Tomato stems</td>
</tr>
<tr>
<td>GRPs</td>
<td>Petunia stems</td>
</tr>
<tr>
<td>GRPs</td>
<td>Petunia stems</td>
</tr>
<tr>
<td>GRPs</td>
<td>Tobacco stems</td>
</tr>
<tr>
<td>GRPs</td>
<td>Tobacco with bean GRP 1.8–GUS transgene</td>
</tr>
<tr>
<td>GRPs</td>
<td>Soybean roots</td>
</tr>
<tr>
<td>GRPs</td>
<td>Bean seeds</td>
</tr>
<tr>
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out of the cut cells of the tissue to the filter membrane. Because the bulk of extensin becomes extensively cross-linked in the wall and insoluble, as discussed below, it does not transfer during the tissue printing process. Additionally, tissue printing of wounded tissue results in the detection of less than expected amounts of extensin, extensin mRNA, and GRP mRNA (A. Butt and A. Showalter, unpublished results). This may be due to degradation, wound healing, or insolubilization. Another problem that must be considered for the extensins in designing and analyzing tissue print RNA or protein blots, in situ hybridizations, and immuncytocrochemical localizations is the possibility that nucleic acid and antibody probes will cross-react with other extensin or extensin-related sequences.

**Intemolecular Interactions and Functions**

The rapid insolubilization of extensins once they are secreted into the wall not only serves as an impediment to extensin research on the protein level, but also represents a major unsolved mystery in terms of how insolubility occurs and, related to this, with what molecules extensins interact. To date, there are several clues but no direct evidence. Extensins may be cross-linked by intermolecular diphenylether linkages between tyrosines, although such isodityrosine cross-links have only been found intramolecularly (Epstein and Lamport, 1984). Other cross-links cannot as yet be ruled out. One study has shown that extensin monomers (i.e., soluble extensin) form a type of extensin oligomer in the presence of a crude cell wall enzyme preparation (Everdeen et al., 1988). Other recent work has shown that the extensin (and PRP) insolubilization is enhanced within just a few minutes of wounding, elicitor treatment, or glutathione treatment (Bradley, 1992). This insolubilization process is hypothesized to be mediated by the release of hydrogen peroxide and catalyzed by a wall peroxidase. This response is thought to be an ultrarapid defense response that serves to further strengthen the cell wall.

Extensin may also be covalently cross-linked to some wall carbohydrate(s). This was suggested years ago (Keegstra et al., 1973); however, X. Qi and A. Mort (personal communication) have only recently found biochemical evidence supporting the existence of an extensin–pectin cross-link.

It is also likely that extensin interacts ionically with pectins. The positively charged lysine and protonated histidine residues of extensin are candidates for ionic interactions with the negatively charged uronic acids of pectins. Such interactions could be regulated by changes in cell wall pH and Ca\(^{2+}\) and thus alter the physicochemical properties of the wall. Lysine residues may also form Schiff base linkages with polysaccharides, and immuncytocrochemical localizations is the possibility that nucleic acid and antibody probes will cross-react with other extensin or extensin-related sequences.

**GLYCINE-RICH PROTEINS**

**Structure**

GRPs represent a relatively newly discovered class of plant proteins that are characterized by their repetitive primary structure, which contains up to 70% glycine arranged in short amino acid repeat units. The first GRP gene was isolated by Condit and Meagher (1986), who used Epstein-Barr virus DNA as a probe to try to isolate oncogenes from petunia. The protein encoded by this gene contains 67% glycine arranged predominantly in Gly-X repeat units, where X is most frequently Gly but can also be Ala or Ser. The observation that this GRP gene encoded a signal peptide sequence, together with protein data showing that a cell wall protein fraction of pumpkin seed coat contained 47% glycine (Varner and Cassab, 1986), suggested that GRPs are cell wall proteins. Keller et al. (1988) subsequently isolated a bean genomic clone containing two linked GRP genes that predicted proteins containing 63% and 58% glycine in which the glycines are arranged predominantly in repeating Gly-X units, similar to those predicted by the petunia GRP gene. Several other groups have also isolated and characterized GRP cDNAs or genes from tomato (Godoy et al., 1990; Showalter et al., 1991), Arabidopsis (de Oliveira et al., 1990; Quigley et al., 1991), petunia (Linhorst et al., 1990), tobacco (van Kan et al., 1988; Obokata et al., 1991), carrot (Sturm, 1992), and Chenopodium rubrum (Kaldenhoff and Richter, 1989). Most of these GRP clones predict one common protein feature, the presence of an amino terminal signal peptide. The idea that these GRPs are localized in the cell wall has subsequently been verified by immunolocalization studies with antibodies against the petunia GRP and one of the bean GRPs (Keller et al., 1988, 1989b; Condit et al., 1990). Because some of the dicot GRP clones apparently lack signal peptides and at least one of them also encodes an RNA binding
GRPs are not limited to dicotyledonous plants or, apparently, to the cell wall. Genes encoding GRPs have been isolated from maize (Gómez et al., 1988) and rice (Mundy and Chua, 1988; Lei and Wu, 1991), and GRP cDNAs have been isolated from maize (Didierjean et al., 1992), sorghum (Crétin and Puigdomènech, 1990), and barley (Rohde et al., 1990). The maize GRP gene encodes a protein that is 37% glycine and consists largely of Gly-GlyTyr-Gly-Gly and Arg-Arg-Glu amino acid repeats. One rice GRP gene encodes a protein that contains only 25% glycine and has no recognizable repeat units (Mundy and Chua, 1988). Interestingly, neither the rice GRP nor the maize GRP includes a signal peptide sequence, making their localization in the cell wall unlikely. In fact, antibodies produced against the rice GRP demonstrated a cytosolic location for this protein. By contrast, another rice GRP was found to encode a signal peptide and a mature protein containing 67% glycine (Lei and Wu, 1991). This rice GRP is clearly homologous to the petunia GRP and to one of the bean GRPs, GRP 1.8, and is thought to reside in the cell wall, not only because it contains a signal peptide but also because the bean GRP 1.8 antibody cross-reacts with an antigen contained in a rice cell wall fraction. The barley GRP cDNA also encodes a putative signal peptide and specifies a unique GRP that has some sequence homology to vertebrate cytokeratins. The sorghum GRP cDNAs appear to be very similar to the maize GRP and do not encode signal peptides. Of as yet unknown functional significance is the observation that the GRPs encoded by both the maize and sorghum GRP clones contain a common RNA binding sequence (Mortenson and Dreyfuss, 1989).

Regulated Expression

As shown in Table 3, dicot GRPs, like extensins, are expressed in response to a variety of developmental and stress conditions (Condit and Meagher, 1987; Keller et al., 1988; van Kan et al., 1998; Condit et al., 1990; de Oliveira et al., 1990; Linthorst et al., 1990; Showalter et al., 1991, 1992). Monocot GRPs are likewise expressed in response to a similar set of conditions (Gómez et al., 1988; Mundy and Chua, 1988; Lei and Wu, 1991; Didierjean et al., 1992).

Dicot GRPs are commonly localized to vascular bundles, particularly to xylem elements, and are clearly associated with cells that are going to be lignified (Table 4). Moreover, these GRPs are usually colocalized with the PRPs (Table 4). In contrast, Gómez et al. (1988) localized the maize GRP mRNA to scutellar epidermal cells surrounding the embryo and to epidermal cells of embryonic leaves.

Intermolecular Interactions and Functions

The picture that has emerged is that there are at least two broad classes of GRPs. One class is found in the cell wall and is developmentally regulated, whereas the second class is found somewhere in the cytoplasm and is regulated by a variety of stress conditions, including abscisic acid and drought stress. Both GRP classes are represented in dicots and in monocots. The cell wall GRPs are most likely structural proteins that presumably have important functions with respect to plant vascular systems and wound healing. Secondary structure predictions for these cell wall GRPs indicate that they exist as β-pleated sheets composed of varying numbers of antiparallel strands as dictated by the particular GRP sequence; such a structure could provide elasticity as well as tensile strength during vascular development. The association of these GRPs with cells destined to become lignified is intriguing, and it has been suggested that they may serve as nucleation sites for such an event, possibly by the catalytic action that their tyrosine residues could have on the oxidative polymerization chain reaction of lignin synthesis (Keller et al., 1989b). The non-cell wall GRPs may play some role in drought tolerance as well as in wound healing. In addition, the presence of a consensus RNA binding site in at least some of these GRPs may indicate still another function.

One of the bean GRPs, GRP 1.8, is apparently insolubilized in the wall (Keller et al., 1989b); whether this will be the case for the other cell wall GRPs is unknown. Also unknown is what could cause this insolubilization. One possibility is that intermolecular isodityrosine linkages could be formed with other GRP molecules or wall proteins. Based upon recent tissue print data, an extensin-GRP cross-link is unlikely given that these wall proteins are not found in abundance in the same cell types. Perhaps more likely is their interaction with PRPs, given their colocalization, or their interaction with lignin, given that GRPs are deposited in tissues destined to be lignified (Ye et al., 1991). With respect to the latter point, it should be noted that tyrosine residues are capable of forming covalent bonds with dehydrogenated coniferyl alcohol in a reaction catalyzed by peroxidase in vitro (Whitmore, 1978).

Extensin clones have been used by Keller et al. (1988), Showalter et al. (1991), and Lei and Wu (1991) to select accidently and serendipitously GRP clones from bean genomic, tomato cDNA, and rice genomic libraries, respectively, due to nucleotide sequence homology between these two protein classes. This homology is not unexpected and arises from the sequence complementarity between stretches of CCX, which encodes proline, and stretches of GGX, which encodes glycine. In other words, the noncoding strand of an extensin gene shares sequence homology to the coding strand of a GRP gene and vice versa. To date, no extensin gene has been found to be transcribed in the opposite orientation to give rise to a GRP mRNA or vice versa. It should also be noted that there are no sequence motifs corresponding to GlyXAsp/Glu in the GRPs (these would be encoded by the complementary sequence giving rise to Ser-HypX repeats of dicot extensins); instead, glycine residues occur in clusters of no apparent pattern (with the exception of the Gly-X motif observed in several GRPs, where X can be any of several possible amino acids including Gly). Similarly, no maize THRGP sequence motif can be encoded by the maize GRP gene. Perhaps a gene encoding an HRGP containing long stretches of hydroxyproline, such
as that found in Volvox but not yet cloned (Mitchell, 1980), will prove to be the most homologous sequence to the GRP genes; only additional study will tell.

PROLINE-RICH PROTEINS

Structure

PRPs represent another relatively newly identified class of plant cell wall proteins of which at least some, and perhaps all, members contain hydroxyproline. There are at least two broad subclasses of PRPs: those that are components of normal plant cell walls (Chen and Varner, 1985a; Hong et al., 1987; 1990; Averyhart-Fullard et al., 1988; Tierney et al., 1988; Datta et al., 1989; Datta and Marcus, 1990; Kleis-San Francisco and Tierney, 1990; Lindstrom and Vodkin, 1991; Sheng et al., 1991) and those that are plant nodulins (i.e., proteins produced in response to infection by nitrogen-fixing bacteria) and constitute part of the nodule cell wall (Franssen et al., 1987, 1989; Dickstein et al., 1988; Scheres et al., 1990; van de Wiel et al., 1990; Govers et al., 1991). The distinction between these two classes may not be clear cut because two pea nodulin PRPs, ENOD12A and ENOD12B, are also expressed in stems and flowers (Scheres et al., 1990; Govers et al., 1991).

All of the PRPs are characterized by the repeating occurrence of Pro-Pro repeats that are contained within a variety of other larger repeat units. For example, many of the normal cell wall PRPs and some nodulin PRPs, such as soybean, alfalfa, and pea ENOD2, are characterized by the presence of the repeating pentapeptide sequence Pro-Pro-X-Y-Lys, where X and Y can be valine, tyrosine, histidine, and glutamic acid. In some cases, the repetitive unit extends an additional proline residue beyond the other two proline residues. Those PRPs that have been isolated and characterized lack carbohydrate or are only lightly glycosylated (Datta et al., 1989) and contain approximately equimolar quantities of proline and hydroxyproline (Averyhart-Fullard et al., 1988; Datta et al., 1989; Kleis-San Francisco and Tierney, 1990). In addition, amino acid sequence analysis has shown that hydroxyproline occurs exclusively in the second position of a soybean cell wall PRP pentapeptide repeat (i.e., Pro-Hyp-X-Y-Lys) (Lindstrom and Vodkin, 1991) and in the second and third positions as well as only in the third position of two hexapeptide repeats found in a gymnosperm PR (i.e., Pro-Hyp-Hyp-X-Y-Lys and Pro-Pro-Hyp-Val-Val-Lys, respectively) (Kielszewski et al., 1992a). It will be interesting to observe the hydroxylation patterns of the nodulin PRPs and determine whether they are consistent with the above motifs and other rules postulated for proline hydroxylation.

José-Estanyol et al. (1992) recently described the cloning and characterization of the first monocot PRP gene. This maize PRP consists of an N-terminal proline-rich domain with numerous Pro-Pro-Tyr-Val and Pro-Pro-Thr-Pro-Arg-Pro-Ser repeats and a C-terminal proline-poor domain that is hydrophobic and contains several Cys residues. This characteristic domain arrangement is similarly exhibited by two dicot PRP sequences, one from bean (Sheng et al., 1991) and the other from tomato (Salts et al., 1991).

Regulated Expression

Regulatory studies indicate that PRPs are involved in various aspects of development, ranging from germination to the early stages of nodulation (reviewed in Showalter and Rumeau, 1990; Hong et al., 1989; Scheres et al., 1990; José-Estanyol et al., 1992). In addition, it has been reported that wounding, endogenous elicitors, fungal elicitor, ethylene, cell culturing, and light can affect PRP gene expression (Tierney et al., 1988; Marcus et al., 1991; Sheng et al., 1991).

Members of the PRP gene family exhibit tissue- and cell-specific patterns of expression (Table 4). As already mentioned, PRPs commonly show a localization pattern similar to that of most dicot GRPs, although some PRP expression occurs in the same cell types as extensins. Recently, Wyatt et al. (1992) used in situ hybridization with gene-specific probes to reveal different tissue-specific expression patterns for three members of the soybean PRP gene family in 4-day-old soybean hypocotyls (Table 4). Indeed, such gene-specific/protein-specific studies are important as they allow for the dissection of the gene/protein family without the complication of cross-hybridization or cross-reactivity to related family members. In the case of PRP nodulins and the maize PRP, other distinct patterns of expression are seen.

Intermolecular Interactions and Functions

It is likely that the PRPs, like the extensins and GRPs, are insolubilized in the wall over time. Indeed, data already exist that support this idea (Datta et al., 1989; Kleis-San Francisco and Tierney, 1990). Moreover, this insolubilization process can occur rapidly in response to stress and may be mediated by the release of hydrogen peroxide and catalyzed by a wall peroxidase (Bradley et al., 1992). The molecular interactions that cause insolubilization, however, are unknown. The relatively high tyrosine content of the PRPs raises the possibility of isodityrosine cross-links between PRP molecules and/or between PRPs and GRPs or extensins, given the above tissue localization data. As with the GRPs, the association of some PRPs with lignification raises the possibility that they are involved in the lignification process (Ye et al., 1991). In addition, given that the PRPs tend to be basic proteins because of their high lysine content and that salt extraction can solubilize some, but not all, soybean PRPs from the wall, it is conceivable that PRPs interact ionically with other wall components such as the acidic pectins, as is also likely for the extensins.

The PRPs may have important roles in normal development and in nodule formation, but their precise functions are unknown. In recent work, peptidyl proline hydroxylase inhibitors were applied to soybean cells in culture, with the result that
salt-extractable PRPs were reduced and cell growth was arrested (Schmidt et al., 1991). A similar study performed several years ago showed that tobacco cells treated with prolyl hydroxylase inhibitors led to the production of monster cells (i.e., cells that continued to grow but did not divide; J. Cooper, personal communication). Clearly, such experiments are interesting and their correlations intriguing; however, two complications should be kept in mind, namely that the inhibitors are not totally specific and may affect general cell metabolism, and that all hydroxyproline-containing proteins will be affected, making it impossible to ascribe a function to any one particular HRGP.

In the case of the PRP nodulins, roles in nodule morphogenesis and in the bacterial infection process are possible. Based on their expression patterns, the ENOD2 class of PRP nodulins from soybean, alfalfa, and pea is hypothesized to have as yet undefined roles in the bacterial infection process and in developmental processes occurring in other plant tissues as well (Scheres et al., 1990).

There is an obvious bit of sequence identity between the PRPs and the extensins (Table 2). This is best demonstrated by comparing the Pro-Pro-Val-Tyr-Lys repeat common to many PRPs with the Ser-Hyp-Pro-Val-Tyr-Lys repeats of tomato P2 extensin and the Ser-Hyp-Thr-Pro-Val-Tyr-Lys repeats of carrot, tobacco, and petunia extensins. Similarity can also be seen between the N-75 (or ENOD2) nodulin sequence Pro-Pro-His-Glu-Lys-Pro-Pro-Pro and a portion of the sugar beet extensin sequence Hyp-Hyp-Thr-Pro-Val-Tyr-Lys repeats of carrot, tobacco, and petunia extensins. Certain PRP sequences also show limited sequence identity to the N-terminal domain of y-zein, which contains Pro-Pro-Pro-His-Leu-Pro-Pro repeats, and to the animal salivary proline-rich proteins (Showalter and Rumeau, 1990).

**SOLANACEOUS LECTINS**

**Structure**

Lectins are carbohydrate binding proteins or glycoproteins of diverse origin. The solanaceous lectins represent a unique class of plant lectins that can be distinguished from other lectins by their restricted occurrence in solanaceous plants, their ability to agglutinate oligomers of N-acetylglucosamine, their predominantly extracellular location, and their unusual amino acid and carbohydrate composition in which hydroxyproline and arabinose are major constituents (reviewed in Allen, 1983; Showalter and Varner, 1989).

Potato tuber lectin (PTL) is the most well-studied member of the solanaceous lectins. It is a glycoprotein with a monomeric molecular weight of 50 kD and consists of 50% carbohydrate and 50% protein by weight (Allen and Neuberger, 1973; Allen et al., 1978; Matsumoto et al., 1983). The carbohydrate moiety consists mainly of arabinose, which is linked to hydroxyproline. These linkages generally take the form of tri- and tetra-arabinosides; in addition, and to a lesser extent, galactose is linked to serine (Allen et al., 1978; Muray and Northcote, 1978; Ashford et al., 1982). The protein moiety is especially rich in four amino acids, hydroxyproline, serine, glycine, and cysteine. In its native form, it is extensively linked by disulfide bridges so that no free sulfhydryl groups are present. Exhaustive pronase digestion of PTL following reductive alkylation releases a 33-kD glycopeptide that is exceptionally rich in serine and hydroxyproline but not in glycine or cysteine (Allen et al., 1978). These data indicate that PTL consists of at least two distinct protein domains; one is rich in serine and hydroxyproline and contains the carbohydrate moiety, and the other(s) is rich in glycine and cysteine. An analogous experiment with the *Datura stramonium* lectin led to an essentially identical conclusion (Desai et al., 1981). The ability of PTL to bind N-acetylglucosamine oligomers is known to be associated with the glycine-cysteine-rich domain(s) (Allen et al., 1978; Ashford et al., 1981), and it is likely that this chitin binding domain will be homologous to that found in other chitin binding proteins such as wheat germ agglutinin, barley lectin, rice lectin, stinging nettle lectin, hevein, potato win1 and 2, poplar win8, bean chitinase, potato chitinase, and tobacco chitinase (reviewed in Chrispeels and Raikhel, 1991). In the case of many of these chitin binding protein genes, the chitin binding domain is fused to an unrelated protein domain.

The serine-hydroxyproline-rich glycopeptide domain of PTL and the other solanaceous lectins bears a striking biochemical resemblance to the extensins. The similarity between these two classes of HRGPs is reflected in their extracellular location, their abundance of serine, hydroxyproline, and arabinose, and in the presence of identical carbohydrate–protein linkages, namely hydroxyproline attached predominantly to tri- and tetra-arabinosides and serine attached to single galactose residues. These similarities probably reflect an interesting evolutionary relationship between these two classes of HRGPs that remains to be elucidated. Indeed, it is likely that the repeating Ser-Hyp2 pentapeptide sequences that are characteristic of most dicot extensins will also characterize the serine-hydroxyproline-rich glycopeptide domain of PTL and the other solanaceous lectins.

**Regulated Expression**

It has been reported that PTL accumulates in potato tubers in response to wounding (Casalongué and Pont Lezica, 1985) and viral infection (Scheggia et al., 1988). It is also reported that PTL agglutinates avirulent, but not virulent, strains of *Pseudomonas solanacearum* (Sequeira and Graham, 1977) and that a hydroxyproline-rich lectin, curiously enough from bean plants, accumulates in bean suspension cultures in response to fungal elicitor (Bolwell, 1987).

Tissue print immunoblots have shown that PTL is deposited in a developmental fashion, with initial deposition occurring
in the cortex and pith regions of the tuber and subsequent distribution occurring uniformly in the whole tuber (Pont-Lezica et al., 1991). Additional tissue immunoblots revealed that the potato lectin is found in both the outer and inner phloem of other potato tissues (R. Pont-Lezica, personal communication). Unfortunately, because the potato lectin antibodies cross-react with extensin, it is impossible to know the contribution being made by each protein in these immunoblots.

**Intermolecular Interactions and Functions**

Because the solanaceous lectins appear to be localized in the cell wall and appear more or less similar to many dicot extensins, these lectins conceivably have the potential to interact with cell wall components. However, the tyrosine and lysine content is considerably less than that of most dicot extensins, making isodityrosine cross-links and ionic interactions with negatively charged wall components unlikely. The glycine-cysteine-rich domain would be available to interact with wall components as well, but it is perhaps more likely that this domain would be used for binding pathogens that have carbohydrate specificity. Oligomers may be important.

**ARABINOGLACTAN PROTEINS**

**Structure**

AGPs are HRGPs that are generally very soluble and highly glycosylated (reviewed in Fincher et al., 1983; Showalter and Varner, 1989). AGPs are widely distributed in plants and typically comprise only 2 to 10% protein by weight. Their molecular weights are extremely heterogeneous, presumably reflecting different extents of glycosylation. AGPs are readily solubilized during tissue extraction with low ionic strength aqueous solutions. Most AGPs are also soluble in saturated ammonium sulfate. These solubility properties have greatly facilitated their isolation and characterization. The protein moiety of AGPs is typically rich in hydroxyproline, serine, alanine, threonine, and glycine and is resistant to proteolysis in its native state, a property that is presumably conferred by extensive glycosylation. AGPs have isoelectric points in the range of pH 2 to 5. The N-terminal sequences of four different AGPs, three from carrot and one from ryegrass, have been determined; all four sequences contain Ala-Hyp repeats (Gleeson et al., 1985). More recently, sequences from five tryptic peptides from deglycosylated ryegrass AGP have also been determined; some of these sequences also contain Ala-Hyp repeats (Gleeson et al., 1999). Pending further investigation, such repeats may become a diagnostic characteristic of AGPs, because they are not known to occur in any of the other cell wall proteins, with the possible exception of the maize HHRGP (Table 2) (Kieliszewski et al., 1992b).

Carbohydrates account for most of the weight of AGPs, and, as their name implies, AGPs contain D-galactose and L-arabinose as the major carbohydrate constituents. Structural analysis has shown that carbohydrate moiety of some AGPs consists of polysaccharide chains with (1→3) linked β-D-galactopyranose residues branched with (1→6) linked β-D-galactopyranose side chains, which are in turn branched with arabinofuranose and other, less abundant monosaccharides (Fincher et al., 1983). Moreover, such moieties appear to be attached regularly to the protein core via β-D-galactopyranose–hydroxyproline linkages (Fincher et al., 1983; Bacic et al., 1997). It is worth noting that carbohydrate moieties similar, if not identical, to those present in AGPs also exist in the cell wall unattached to protein; these are the arabinogalactans, which have been reviewed elsewhere (Clarke et al., 1979).

Conclusive cellular localization of many AGPs is difficult because of their extreme solubility. It is clear, however, that AGPs are often found as constituents of the extracellular milieu. Cells in suspension culture from various tissue sources secrete AGPs into the culture medium; also, certain specialized cells, such as the stylar canal cells and some secretory cells that produce gummy exudates, secrete especially large quantities of AGPs into the cell wall compartment (Clarke et al., 1979). In addition, some AGPs are associated with the plasma membrane (Pennell et al., 1989; Norman et al., 1990). Yariv's antigen, a β-D-glucosyl artificial carbohydrate antigen, specifically precipitates most AGPs as a red–orange complex, greatly aiding the cellular and tissue localization studies, even though the interaction between Yariv's antigen and AGPs is not well understood (Fincher et al., 1983). This interaction also serves to illustrate the point that most AGPs are "β-lectins," with a broad binding specificity directed toward β-D-glycopyranosyl linkages.

Electrophoretic separation of AGPs in the presence of Yariv's antigen shows that AGPs are expressed in a tissue-specific manner and that a given plant tissue can contain more than one kind of AGP (van Holst and Clark, 1986). Whether these AGPs differ with respect to their protein moieties, carbohydrate moieties, or both is not yet known. In this context, Norman et al. (1990) have shown that a set of tobacco plasma membrane AGPs are produced by differential glycosylation of a 50-kD core protein.

Circular dichrometry shows that ~30% of the protein moiety of an AGP is in a polyproline II helix (van Holst and Fincher, 1984). Little is known about the conformation of the rest of the
protein or the carbohydrate chains. One model, based on the above information and the observation that some AGPs have low viscosities, predicts that carbohydrate moieties with ovoid or spheroidal shapes are attached at several positions on a core protein (Fincher et al., 1983).

Regulated Expression and Functions

No function(s) for AGPs has been established unequivocally. Based on their predominantly extracellular location and their various biochemical and physical properties, AGPs have been proposed to act as glues, lubricants, and humectants (Fincher et al., 1983). Their general abundance in the middle lamellae of the wall and in the styles of angiosperms and in the medulla of root nodules (Cassab, 1986) makes them likely candidates for functioning in cell-cell recognition. There is also some indication that AGPs accumulate in response to wounding (Fincher et al., 1983). Consequently, a role for AGPs in wound healing or plant defense is a possibility that requires further investigation.

Evidence for developmental regulation of plasma membrane AGPs in flowers, embryos, and roots has recently been obtained (Pennell and Roberts, 1990; Knox et al., 1991; Pennell et al., 1991). Thus, a role for these AGPs in floral histogenesis and differentiation is possible. Such a role may involve cell-cell interactions in which these plasma membrane AGPs act as cell adhesion molecules analogous to mammalian substrate adhesion molecules and capable of binding as yet undefined cell wall ligands (Pennell et al., 1991). Herman and Lamb (1992) have shown that plasma membrane AGPs are not only localized to tobacco leaf plasma membranes but also to intravacular multivesicular bodies. This observation has led them to propose that plasma membrane AGPs are involved with an endocytotic pathway for vacuolar-mediated disposal of the periplasmic matrix.

FUTURE DIRECTIONS AND QUESTIONS

Relatively straightforward structural and regulatory studies have now been done on numerous examples of the five major cell wall proteins. Such characterization work will surely continue and new family members will be isolated with perhaps a few surprises along the way. Regulatory studies of the wall protein genes will undoubtedly go the route of so many other genes, and cis- and trans-acting factors will be identified by now fairly routine molecular biology technology. Cloning of the as yet uncloned solanaceous lectins and AGPs should soon follow. Furthermore, it is almost inevitable that new wall proteins and their genes will be identified and characterized.

More difficult questions of function and molecular interactions are ahead, however. To ascertain the function of these proteins, several approaches are possible. First, knowing where the proteins are located in the plant (i.e., in which organs and tissues) will be an important step in ascribing a function to these proteins. Such studies alone will not provide the functional answer, but they will provide critical clues, and any functional assignment will ultimately have to be consistent with the results of the localization studies. The most direct and unambiguous way to address function is to produce cell wall protein mutants and examine phenotypic alterations of mutant plants. Mutants could be produced with antisense RNA technology, with the aid of ribozymes, or by mutagenesis followed by screening with antibodies to the various wall proteins. Some of this work has begun in the labs of James Cooper, Andrew Staehelin, and Keith Roberts, but no clearcut picture has emerged. Indeed, these are exciting but difficult experiments because many questions concerning them can be raised. For what function does one look? Is one looking at the right place and time? Is one looking under the correct conditions to see an altered phenotype? Will other members of the gene family or other wall proteins compensate for the loss of one wall protein?

As for determining how the cell wall proteins interact in the wall, several approaches are possible: labeling the cells and characterizing proteolysis products from the cell wall, adding labeled synthetic peptides to cell cultures and testing for insolubilization and subsequent characterization of the cross-links, producing tagged chimeric proteins (i.e., part marker protein, part cell wall peptide motif) in transgenic plants and examining insolubilization and cross-links, purifying cell wall enzyme preparations capable of cross-linking purified cell wall protein monomers in vivo, and examining cell wall protein connections in plants via solid state nuclear magnetic resonance. Clearly, these are for the most part difficult biochemical experiments, and their interpretations may be equally difficult, particularly with respect to possible functional implications.

Our current state of knowledge has allowed for some evolutionary speculation, but more sequences are needed, particularly from more primitive plants, to see these evolutionary connections (or artifacts) more clearly. Is it really possible that all of the HRGPs, and even the GRPs, represent products of one giant supergene family? Indeed, some intriguing evolutionary studies lie ahead.

Finally, it would be of great use to select a model system for plant cell wall research. Arabidopsis is one logical choice, particularly because it is so amenable to mutagenesis studies to address functional questions, but others are clearly possible. Examining all of the plant cell wall components, including all of the major structural wall proteins, together in one system would allow for a more integrated approach to elucidate cell wall structure, intermolecular interactions, and ultimately cell wall function.

ACKNOWLEDGMENTS

The preparation of this review was made possible by an Academic Challenge Grant from the state of Ohio and by United States Department
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Received January 27, 1992; accepted November 4, 1992.


Structure and function of plant cell wall proteins.
A M Showalter
Plant Cell 1993;5;9-23
DOI 10.1105/tpc.5.1.9

This information is current as of November 4, 2017

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