Molecular Remodeling of Photosystem II during State Transitions in *Chlamydomonas reinhardtii* (Minagawa and Takahashi, 2004)

Masakazu Iwai, Yuichiro Takahashi, and Jun Minagawa

A Institute of Low Temperature Science, Hokkaido University, N19 W8, Sapporo 060-0819, Japan  
B Department of Biology, Faculty of Science, Okayama University, Okayama 700-8530, Japan

State transitions, or the redistribution of light-harvesting complex II (LHClII) proteins between photosystem I (PSI) and photosystem II (PSII), balance the light-harvesting capacity of the two photosystems to optimize the efficiency of photosynthesis. Studies on the migration of LHClII proteins have focused primarily on their reassociation with PSI, but the molecular details of their dissociation from PSII have not been clear. Here, we compare the polypeptide composition, supramolecular organization, and phosphorylation of PSII complexes under PSI- and PSII-favoring conditions (State 1 and State 2, respectively). Three PSII fractions, a PSII core complex, a PSII supercomplex, and a multimer of PSII supercomplex or PSII megacomplex, were obtained from a transformant of the green alga *Chlamydomonas reinhardtii* carrying a Histagged CP47. Gel filtration and single particles on electron micrographs showed that the megacomplex was predominant in State 1, whereas the core complex was predominant in State 2, indicating that LHClIs are dissociated from PSII upon state transition. Moreover, in State 2, strongly phosphorylated LHCII type I was found in the supercomplex but not in the megacomplex. Phosphorylated minor LHClIs (CP26 and CP29) were found only in the unbound form. The PSII subunits were most phosphorylated in the core complex. Based on these observations, we propose a model for PSII remodeling during state transitions, which involves division of the megacomplex into supercomplexes, triggered by phosphorylation of LHCII type I, followed by LHCII undocking from the supercomplex, triggered by phosphorylation of minor LHClIs and PSII core subunits.

INTRODUCTION

Photosystem II (PSII) is a large multisubunit pigment protein complex that uses light energy to oxidize water and reduce plastoquinone. In PSII, light energy is captured by the peripheral antenna and is transferred to the core complex, where it is trapped. In green plants, the peripheral antennas are composed of major trimeric and minor monomeric light-harvesting complex II (LHClII) proteins (Dekker and Boekema, 2005). In spinach, another supercomplex, C2S2M2 supercomplex, has been reported, where two moderately bound LHCII trimers along with CP24 are associated with the C2S2 supercomplex (Boekema et al., 1998).

Photosystem I (PSI) has a broad absorption peak in the far-red region as well as peaks in the blue and red regions, whereas PSII has peaks in the blue and red but not in the far-red region. Thus, an imbalance of energy distribution between the two photosystems tends to occur in natural environments, where light quality and quantity fluctuate with time (Bollaire et al., 2005; Tikkanen et al., 2006). Under such conditions, state transitions occur to balance the light-harvesting capacity of the two photosystems (Bonaventura and Myers, 1969; Murata, 1969). When the plastoquinone pool gets reduced (Allen et al., 1981), a protein kinase is activated through the cytochrome bf complex (Wollman and Lemarie, 1988; Vener et al., 1997). This kinase phosphorylates the LHCI that is bound to PSII in the appressed region of the thylakoids (Bennett, 1977). The phosphorylation of LHCI then leads to the lateral migration of LHCI to PSI in the unappressed region (Andersson et al., 1982), where it acts as the peripheral antenna for PSI (State 2). Oxidation of the plastoquinone pool induces the opposite effect, regenerating State 1 (Bennett, 1980).
The association of LHClI proteins with PSI has been observed indirectly as changes in the quantum yield of PSI and PSII during state transitions (Samson and Bruce, 1995). Phosphorylated LHClI polypeptides were detected in the unappressed region of thylakoid membranes where PSI is preferentially located (Bassi et al., 1988). However, only recently have there been several lines of direct biochemical evidence shown the binding of LHClI polypeptides to PSI (Pesaresi et al., 2002; Zhang and Scheller, 2004; Takahashi et al., 2006). Such an association was first observed in an Arabidopsis psea1-1 mutant where a fraction of LHClI was associated with PSI when the mutant plants were exposed to low-light (State 2 favoring) conditions, giving rise to a high molecular mass protein-pigment complex (Pesaresi et al., 2002). This large complex, however, seemed to be an aggregated product since the mutant did not show state transitions, likely due to a low level of PsAh. The next attempt was through cross-linking of the mobile LHClI proteins with the PSI-LHClI supercomplex in Arabidopsis (Zhang and Scheller, 2004). Major LHClI proteins, including Lhcb1 and -2, were cross-linked to PsAh, I, and L subunits in State 2 than in State 1. A further report was provided from a study on C. reinhardtii, where the PSI-LHClI/II supercomplex isolated from State 2 cells contained three LHClI polypeptides, CP26, CP29, and LhcbM5, suggesting a pivotal role for the minor monomeric LHClI in state transitions (Takahashi et al., 2006). Mobile LHClI proteins were also observed in single particle images of the PSI-LHClI/II supercomplexes. Boekema and colleagues (Kouril et al., 2005) reported a large density along the side of PsAh/L/A/K in electron micrographs of Arabidopsis PSI, which they assigned to the LHClI trimer, and Barber and colleagues located a smaller density near PsAh in C. reinhardtii, which they assigned to CP29 (Kargul et al., 2005).

Although much has been learned about the molecular details of state transitions on the PSI side, those on the PSII side remain elusive. Which LHClI polypeptides are dissociated from PSII? What is the dissociation order? Does the dissociation cause any effects on PSII macroorganization? Notably, the roles of phosphorylated LHClI have been obscure. It was originally proposed that the phosphorylation of LHClI induces conformational changes, leading to its dissociation from PSII (Nilsson et al., 2006). Such an association was first observed in an Arabidopsis psae1-1 mutant where a fraction of LHClI was associated with PSI when the mutant plants were exposed to low-light (State 2 favoring) conditions, giving rise to a high molecular mass protein-pigment complex (Pesaresi et al., 2002). This large complex, however, seemed to be an aggregated product since the mutant did not show state transitions, likely due to a low level of PsAh. The next attempt was through cross-linking of the mobile LHClI proteins with the PSI-LHClI supercomplex in Arabidopsis (Zhang and Scheller, 2004). Major LHClI proteins, including Lhcb1 and -2, were cross-linked to PsAh, I, and L subunits in State 2 than in State 1. A further report was provided from a study on C. reinhardtii, where the PSI-LHClI/II supercomplex isolated from State 2 cells contained three LHClI polypeptides, CP26, CP29, and LhcbM5, suggesting a pivotal role for the minor monomeric LHClI in state transitions (Takahashi et al., 2006). Mobile LHClI proteins were also observed in single particle images of the PSI-LHClI/II supercomplexes. Boekema and colleagues (Kouril et al., 2005) reported a large density along the side of PsAh/L/A/K in electron micrographs of Arabidopsis PSI, which they assigned to the LHClI trimer, and Barber and colleagues located a smaller density near PsAh in C. reinhardtii, which they assigned to CP29 (Kargul et al., 2005).

RESULTS

Isolation of PSII Complexes in States 1 and 2

Since relatively few attempts have been reported for purifying the Chlamydomonas PSI-LHClI supercomplex, we sought to establish a streamlined protocol to obtain discrete and intact PSII complexes retaining peripheral antennas. Thylakoids prepared from a genetically modified C. reinhardtii strain, PsbB-His, in which a 6x His-tag was fused to the C terminus of CP47 (Suzuki et al., 2003), were subjected to nickel affinity chromatography. A highly active PSII core complex with no attached LHClIs was previously purified from this strain using β-DM as a detergent (Suzuki et al., 2003). To solubilize PSII particles that retain peripheral LHClIs, we first tested a β-DM epimer n-dodecyl-α-O-maltoside, which has been reported to partially solubilize spinach thylakoids and allow the purification of PSII-LHClI supercomplexes (Boekema et al., 1998). However, n-dodecyl-α-O-maltoside yielded similar results to β-DM in C. reinhardtii (data not shown). We then tested another mild detergent, TM, which has the same head group as β-DM but with one unit longer aliphatic chain than β-DM.

C. reinhardtii cells were grown under low-light conditions (~20 μmol photons m⁻² s⁻¹) until the early exponential phase, and the culture was split into two: one for preparing thylakoids in State 1 and another for preparing them in State 2. Induction of state transitions was done by manipulating the redox state of the plastoquinone pool as described previously (Bulté et al., 1990). Cells were treated with a PSII inhibitor DCMU for 20 min under low light to oxidize the plastoquinones, which locked them in State 1. State 2–locked cells were obtained by treatment with the uncoupler carbonyl cyanide p-(trifluoromethoxy)phenylhydrazone (FCCP) for 20 min under low light to reduce the plastoquinone pool. Fluorescence emission spectra at 77K of cells in State 1 showed that the peak at 689 nm (F₆₈₉) originating from PSII, was higher than the peak at 718 nm (F₇₁₈) from PSI (Figure 1,
State 1). On the other hand, cells locked in State 2 exhibited the opposite trend, as $F_{718}$ was higher than $F_{689}$ (Figure 1, State 2). These results confirmed that the energy distribution between the two photosystems was altered and the cells were indeed locked in the designated state. The results of these treatments are consistent with previous reports (Bulte et al., 1990; Finazzi et al., 1999).

Thylakoid membranes isolated from States 1 and 2 locked cells were solubilized by TM and loaded onto a nickel affinity column. For comparison, we also isolated PSII cores using β-DM solubilization according to the procedure described by Suzuki et al. (2003). Protein complexes carrying His-tagged CP47 were bound to the column and eluted with 250 mM imidazole. Immunoblot analysis confirmed that PSI and PSII subunits were clearly separated in the flow-through and the eluate fractions, respectively (see Supplemental Figure 1 online), indicating that nickel affinity chromatography could be used to purify PSII particles. Each eluate was analyzed by SDS-PAGE and immunoblotting (Figures 2A and 2B) to confirm that the eluted complexes harbor PSII core subunits, including D1, D2, CP43, CP47, PsbO, PsbP, and PsbQ. In addition, the TM-solubilized protein complexes eluted from the column contained minor LHCIIIs (CP26 and CP29) and major LHCIIIs type I, III, and IV, suggesting that these eluates contain PSII complexes retaining the peripheral antennas (Figures 2A and 2B). The retained LHCIIIs were estimated to represent 43 and 16% of LHCIIIs in the State 1 and 2 samples, respectively, by immunoblotting using an anti-LHCII type I antibody. The LHCIIIs recovered in the flow-through are most likely not, or only marginally, bound to PSII.

The Three PSII Fractions and Their Relative Abundance in States 1 and 2

Comparing the amount of bound LHCIIIs, it is clear that the PSII complexes in State 1 bind more LHCIIIs than in State 2 (Figure 2B). This is consistent with the existing model, where mobile LHCIIIs migrate from PSII to PSI in the transition from State 1 to 2, thereby decreasing the PSII antenna size (see, for instance, Allen and Forsberg, 2001). The decreased amount of LHCIIIs associated with PSII complexes in State 2 (Figures 2A and 2B) suggests that the PSII-LHCII supercomplex has a decreased molecular mass in State 2 compared with State 1. We thus performed gel filtration to analyze the molecular mass of the protein complexes isolated by nickel affinity chromatography. The gel filtration chromatogram showed at least three peaks (or shoulders) at 36.6 (Fraction I), 39.7 (Fraction II), and 43.5 min (Fraction III) retention time, which corresponds to a molecular mass of $2000$, $900$ to $1000$, and $300$ to $400$ kD, respectively (Figure 3). SDS-PAGE and immunoblot analysis confirmed that all three fractions contained PSII core subunits, including D1, CP47, and PsbO (see Supplemental Figures 2 and 3 online). In State 1, the PSII subunits (D1, CP47, and PsbO) were more abundant in Fraction I, whereas in State 2 they were more abundant in Fraction III (see Supplemental Table 1 online). Since

---

**Figure 1.** The 77K Steady State Fluorescence Emission Spectra of *C. reinhardtii* Cells Locked in States 1 and 2.

*C. reinhardtii* cells grown under 20 μmol photons m⁻² s⁻¹ were treated with 10 μM DCMU or 5 μM FCCP for 20 min to induce to State 1 or State 2, respectively. Excitation was given at 440 nm. Spectra were normalized to the 699-nm (CP47) emission peak (Lin and Knox, 1991).
PSII complexes isolated with nickel affinity chromatography were separated by gel filtration on a Superdex 200 PC 3.2/30 column. Three major fractions are designated as I to III. The elution profile of the purified PSII core is shown as a dashed line. Traces are shown for 280-nm absorption factions are designated as I to III. The elution profile of the purified PSII complexes isolated with nickel affinity chromatography were separated by gel filtration on a Superdex 200 PC 3.2/30 column. Three major fractions are designated as I to III. The elution profile of the purified PSII core is shown as a dashed line. Traces are shown for 280-nm absorption.

**Table 1.** Characterization of the Three PSII Fractions Obtained by Ni-Affinity Chromatography and Gel Filtration

<table>
<thead>
<tr>
<th>PSII Fractions</th>
<th>I</th>
<th>II</th>
<th>III</th>
<th>Core</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chlorophyll a/b (w/w)%</td>
<td>3.22 ± 0.37</td>
<td>3.36 ± 0.24</td>
<td>4.30 ± 0.75</td>
<td>4.27 ± 0.13</td>
</tr>
<tr>
<td>Neoxanthin/β-carotene (mol/mol)</td>
<td>0.26 ± 0.13</td>
<td>0.25 ± 0.13</td>
<td>0.13 ± 0.03</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>ε&lt;sub&gt;PSII&lt;/sub&gt; (Å²/quantum)</td>
<td>366 ± 20</td>
<td>350 ± 26</td>
<td>163 ± 88</td>
<td>ND</td>
</tr>
</tbody>
</table>

*a* PSII fractions were isolated from State 2–locked samples. Values shown are means of more than three measurements ± SD.

*b* Chlorophyll concentrations were determined as described by Niyogi et al. (1997).

The relative abundance of Fractions I to III was dramatically changed during state transitions. In State 1, Fraction I was predominant, whereas in State 2, Fraction III was more pronounced (Figure 3). It is thus inferred from these results that there is more LHCII-containing PSII (Fractions I and II) in State 1 and more LHCII-less PSII (Fraction III) in State 2, which is most likely due to the removal of LHCII from the PSII-LHCII supercomplex upon transition from States 1 to 2.

**Figure 3.** Gel Filtration Elution Profile of PSII Particles in States 1 and 2 Isolated with Nickel Affinity Chromatography.

PSII complexes isolated with nickel affinity chromatography were separated by gel filtration on a Superdex 200 PC 3.2/30 column. Three major fractions are designated as I to III. The elution profile of the purified PSII core is shown as a dashed line. Traces are shown for 280-nm absorption. Thick line, State 1; thin line, State 2.

LHCII Polypeptide Composition in the Three PSI Fractions

The composition of LHCII polypeptides in each gel filtration fraction was determined using immunoblotting. Both Fractions I and II contained all of the minor and major LHCII genes except for LHCII type II (Figure 4A). The relative amount of each LHCII to the PSII core (D1 protein) was essentially the same in the two fractions (Figure 4B). In Fraction III, however, the levels of bound LHCII were significantly reduced, confirming that this fraction is mainly composed of PSII complexes that lost most of their LHCII (Figures 4A and 4B).

The major LHCII type I and III in *C. reinhardtii* are encoded by five (LhcbM3, -M4, -M6, -M8, and -M9) and two (LhcbM2 and -M7) duplicated genes, respectively, while both LHCII types II and IV are encoded by single genes, LhcbM5 and -M1, respectively (Minagawa and Takahashi, 2004). To further determine which LHCII gene products are associated with PSII in these fractions, we conducted two-dimensional gel electrophoresis (2-DE). The first dimension was isoelectric focusing, which uncovered distinct LHCII spots because hydrophobic PSII core subunits did not penetrate the gel. Several spots that have been previously reported as LHCII (Stauber et al., 2003) appeared in the region between 20 and 42 kD and pH 4.0 and 5.0 on the 2-DE gels (Figure 5). Polypeptides in these spots were then identified by tandem mass spectrometry (MS/MS) analysis after in-gel trypsin digestion (see Supplemental Table 2 online). All of the minor and major LHCII genes except for LhcBM5 and -M8 were detected in Fraction I (Figure 5). Spot 3, corresponding to CP29, was hardly visualized by silver staining in Figure 5 but was clearly detected by immunoblotting (see Supplemental Figure 5 online) and was confirmed by MS/MS (see Supplemental Table 2 online). The same LHCII polypeptides were identified...
in Fraction II, but they were markedly shifted toward the acidic side (see spots 4 and 5, corresponding to LhcbM3/4/6 and LhcbM4/6/9, respectively in Figure 5, panel II). These shifts could be due to the phosphorylation of these proteins as indicated previously (Hippler et al., 2001). In Fraction III, only a trace amount of LHCII type III (LhcbM2/7) was detected in the 2-DE gel (Figure 5, panel III). These results further support the conclusion that Fractions I and II contain PSII-LHCII supercomplexes with similar polypeptide compositions, and Fraction III contains PSII cores and a small amount of PSII core with LHCII type III. It is of note that among the four types of major LHCIIls in C. reinhardtii, LHCII type II was barely detected in any of the PSII-LHCII supercomplexes in either State 1 or 2; however, it has been reported to be associated with PSI in State 2 (Takahashi et al., 2006). LHCII type II is thus more likely either to be bound at the outer ridge of the peripheral antenna for PSII in State 1, such that it is removed by TM solubilization, or to exist as an unbound form in State 1 and associate with PSI in State 2.

Supramolecular Organization of the Three PSII Particles

According to single-particle image analysis of PSII-LHCII supercomplexes in higher plants, the C_{2}S_{2}M_{2} supercomplex is the most predominant (Dekker and Boekema, 2005). However, the largest PSII-LHCII supercomplex in C. reinhardtii has been reported to be the C_{2}S_{2} supercomplex, which might be due to the lack of CP24 in this organism (Nield et al., 2004; Dekker and Boekema, 2005). The PSII-LHCII supercomplex in Fraction II is likely a C_{2}S_{2} supercomplex, based on the estimated molecular mass and the polypeptide composition (Boekema et al., 1995): two major LHCII trimers, two copies of CP26 and CP29, and one PSII core dimer. The largest PSII particle with molecular mass of \( \approx 2000 \text{ kD} \) in Fraction I could be a multimer of the C_{2}S_{2} supercomplex, since the polypeptide composition (Figure 5), relative amount of LHCIIls to D1 (Figure 4B), chlorophyll a/b ratio,
neoxanthin/β-carotene ratio, and functional absorption cross section (Table 1) of PSII particles in Fraction I are all similar to those in Fraction II.

To investigate the supramolecular organization of the PSII particles in the three gel filtration fractions, negatively stained single particles were observed via electron microscopy. Figure 6 shows a distribution of the top-down views of PSII particles in Fractions I to III. Large (33.0 × 33.0 nm, blue), medium (16.5 × 33.0 nm, green), and small particles (11.0 × 20.5 nm, orange) are predominant in Fractions I, II, and III, respectively (Figure 6; see Supplemental Figure 4 online). These sizes correspond to two C₂S₂ supercomplexes (Nield et al., 2000), one C₂S₂ supercomplex (Nield et al., 2000), and one PSII core dimer (Barber, 2006), respectively. Dekker and Boekema (2005) showed that several types of dimerized PSII-LHCII supercomplexes, so-called megacomplexes, are formed through the lateral association of supercomplexes. Although most of the megacomplexes reported so far in higher plants are dimers of C₂S₂M₂ supercomplexes, the dimerization of C₂S₂ supercomplexes was also suggested in C. reinhardtii (Dekker and Boekema, 2005). Based on these results, it is thus inferred that the predominant PSII particles with molecular masses of ≈2000 kD, 800 to 900 kD, and 300 to 400 kD in Fractions I, II, and III are (C₂S₂)₂ PSII-LHCII megacomplexes, C₂S₂ PSII-LHCII supercomplexes, and PSII core dimers, respectively.

Phosphorylations Involved in the Detachment of LHCII

Phosphorylated polypeptides in the three gel filtration fractions were determined by immunoblotting using an antiphosphothreonine antibody (Figure 7). The obtained results demonstrate that PSII proteins in State 1 were mostly not phosphorylated, whereas PSII-bound LHCII type I and the core subunits CP43 and D2 were strongly phosphorylated in State 2. This clearly indicates that phosphorylation by itself, at least of LHCII type I, does not readily induce detachment of LHCIIIs from PSII. Intriguingly, LHCII type I was much more phosphorylated in Fraction II than in Fraction I in State 2 (Figure 7, Table 2). This implies a possibility that the phosphorylation of LHCII type I triggers the division of the PSII-LHCII megacomplex in Fraction I into the PSII-LHCII supercomplexes in Fraction II. The levels of phosphorylation in the PSII core subunits, CP43 and D2, were highest in the core complex and lowest in the megacomplex (Table 2). Thus, the phosphorylation of the PSII core subunits occurs in a different manner from that of LHCII type I, suggesting a distinct mechanism for the regulation of the kinases for LHCII polypeptides and PSII core subunits.

We also characterized the unbound form of LHCII polypeptides to get further insights on their dissociation from PSII. Free LHCIIIs in flow-through fractions obtained after nickel affinity chromatography were subjected to 2-DE analysis (Figure 8). The compositions of unbound LHCIIIs were nearly the same in State 1 and 2 (Figures 8A and 8B; see Supplemental Figure 5 and Supplemental Table 3 online); however, there were clear differences in their levels of phosphorylation. Spots corresponding to free CP26, CP29, and LHCII type I (LhcbM3, -4, and -6) were more phosphorylated in State 2 (Figures 8C and 8D). Although the free CP26 was also phosphorylated in State 1 (spot 11), possibly more amino acid residues in CP26 were phosphorylated in State 2 (spot 10). In Figure 7, CP26 and CP29 associated with PSII were shown to be nonphosphorylated. Therefore, phosphorylations of CP26 and CP29 most likely lead to their dissociation from PSII. Considering the location of the minor LHCIIIs bordering major LHCII trimers and the PSII core (Harrer et al.,...
we infer that the phosphorylation of minor LHCIIs induces the detachment of all LHCIIs, including major LHCIIs, from PSII. Interestingly, LHCII type III (LhcbM2 and -7) and type IV (LhcbM1) in the flow-through fractions were phosphorylated irrespective of the state (Figures 8C and 8D), whereas those associated with PSII were not (Figure 7). Therefore, unlike LHCII type I, once LHCII type III and IV are phosphorylated they will be undocked from PSII no matter the state.

Lastly, we examined the kinetics of phosphorylation for the most conspicuously phosphorylated proteins of LHCII and PSII in the course of transition from State 1 to 2. Whole cell samples, taken every 2 min under blue light illumination (light 2), were subjected to immunoblotting using an antiphosphothreonine antibody (Figure 9). While the phosphorylation level of LHCII type I was immediately elevated (followed by a slow decline), that of CP29 was gradually increased, but similarly dephosphorylated after 6 min. The phosphorylation of the core subunit CP43 was even slower and reached a plateau after 6 min. The distinct phosphorylation kinetics of the three polypeptides suggests their distinct roles in state transitions. Since Fraction II is the main fraction harboring the phosphorylated LHCII type I (Figure 7), its population could be increased in the initial phase of transition from State 1 to 2. This in turn suggests that the division of the megacomplex occurs first with the LHCII type I phosphorylation and then the other phosphorylations trigger the undocking of LHCIIs from PSII, until just the PSII core complex remains.

**DISCUSSION**

**The Two-Step Transition from State 1 to State 2: Megacomplex to Supercomplex, and Supercomplex to Core Complex**

In this study, PSII particles from State 1– and State 2–locked thylakoids were characterized biochemically and structurally. The profile of the gel filtration and single particle images of the electron micrographs demonstrated that megacomplexes, which are presumably multimers of PSII-LHCII supercomplexes, are predominant in State 1, whereas PSII core complexes with much less LHCII are predominant in State 2. We also identified an intermediate state of transition, the PSII-LHCII supercomplex monomer, which has the same polypeptide and pigment compositions as the megacomplex. We propose that the transition from State 1 to 2 proceeds in two steps on the PSII side: first, the PSII megacomplex is divided into supercomplexes and then LHCIIs (both major and minor) dissociate from these supercomplexes.

**Role of Phosphorylated LHCII**

It is well accepted that LHCII phosphorylation is crucial for the migration of LHCIIs from PSII to PSI (Depe&ége et al., 2003). To give insights into the molecular details of the undocking process of

| Table 2. Relative Phosphorylation Levels of LHCII Type I, CP43, and D2 in the PSII Fractions during State 2 |
|---------------------------------|-----|-----|-----|
| **PSII Fractions**              | I   | II  | III |
| LHCII type I (%)                | 100 | 203 ± 33 | 97 ± 30 |
| CP43 (%)                       | 100 | 142 ± 19 | 160 ± 32 |
| D2 (%)                         | 100 | 159 ± 35 | 172 ± 21 |

* a Phosphorylation levels are normalized to the quantity of D1 protein and quantified by ImageJ. Values shown are means of three measurements ± SD normalized to the value for Fraction I.
LHCII proteins both bound and unbound to PSII was thoroughly characterized. The results suggest distinct roles for each phosphorylated protein: (1) LHCII type I (LhcbM3, -4, -6, and -9), (2) minor LHCII (CP26 and CP29), (3) PSII core subunits (CP43 and D2), and (4) LHCII type III (LhcbM2 and -7) and type IV (LhcbM1).

(1) LHCII Type I (LhcbM3, -4, -6, and -9)

It has been hypothesized that phosphorylated major LHCII proteins do not necessarily dissociate themselves from PSII (Lunde et al., 2000). It is clear that LHCII type I remains associated with PSII upon phosphorylation. Surprisingly, the phosphorylation of LHCII type I is induced earlier in the course of state transition, and it is mostly localized in the supercomplex, not in the megacomplex. This suggests that the phosphorylation of LHCII type I likely causes division of the megacomplex. As has been previously described by Dekker and Boekema (2005), the two CysSrz supercomplexes constituting the megacomplex are bridged by an LHCII trimer in C. reinhardtii. Therefore, the phosphorylation of the most abundant trimeric LHCII, type I, may induce the division of the megacomplex due to its altered conformation (Sprang et al., 1988; Nilsson et al., 1997).

(2) Minor Monomeric LHCII (CP26 and CP29)

Our previous work indicates that the minor monomeric LHCII proteins are shuttled to PSI, thereby acting as a linker between PSI and major trimeric LHCII during a transition to State 2 (Takahashi et al., 2006). The minor LHCII proteins, especially CP29, are indeed strongly phosphorylated in State 2 (Kargul et al., 2005; Takahashi et al., 2006; Turkina et al., 2006). This study provides further evidence for the pivotal role of the minor LHCII proteins. While both CP26 and CP29 associated with the megacomplex and supercomplex were not phosphorylated, those of the free form were phosphorylated (Figure 8D). Thus, only phosphorylated minor LHCII proteins are dissociated from PSI. Since the minor monomeric LHCII proteins are bordering major LHCII trimers and the PSII core (Harrier et al., 1998; Yakushevska et al., 2003), any LHCII proteins associated with PSII will be forced to leave when the minor LHCII proteins are dissociated. Thus, it is inferred that phosphorylation of CP26 and CP29 triggers undocking of the entire peripheral antenna during state 2 transition.

Recently, Turkina et al. (2006) reported the exact phosphorylated amino acid residues on CP29 in C. reinhardtii cells under State 2 as well as under high-light conditions. Transfer of the cells from State 1 to 2 conditions increases the number of phosphorylated residues in CP29 from two to four, whereas high-light treatment causes seven phosphorylated residues. Based on the mapping of those phosphorylated residues at the interface of PSII core and the peripheral antenna proteins, they proposed that the hyperphosphorylation of CP29 causes uncoupling of itself and other LHCII proteins from PSII in cells under State 2 or high-light stress. Our proposal, based on this study, agrees with their report.

(3) PSII Core Subunits (CP43 and D2 Protein)

We demonstrated that CP43 and D2 protein are phosphorylated under State 2 conditions, but the distribution of phosphorylated core proteins among the megacomplex, supercomplex, and core complex, as well as the phosphorylation kinetics, is completely different from that of LHCII. These results are compatible with previous findings that the phosphorylation of PSII core subunits and LHCII proteins were both under the redox control by the plastquinone pool and that only LHCII phosphorylation depends on cytochrome b6 complex (Bennett, 1991). Two distinct kinases, STN7 and STN8, were assigned as the kinases for LHCII and PSII core subunits, respectively (Bellaire et al., 2005; Bonardi et al., 2005).

Since the two phosphorylation events (PSII core subunits and LHCII) occur simultaneously during the transition to State 2, we must emphasize that the two processes are potentially interlinked. Although it is controversial (Andronis et al., 1998; Fleischmann and Rochaix, 1999, Bonardi et al., 2005), PSII core subunit phosphorylation has been suggested to act as a protection mechanism from proteolytic degradation of photodamaged PSII complex under high-light conditions (Koivuniemi et al., 1995; Kruse et al., 1997; Turkina et al., 2006). Since cells were grown in low light in this study (20 μmol photons m−2 s−1), the core subunits are not under the high-light stress. However, the bare PSII core complex (after dissociation of LHCII proteins during state 2 transition) is under high-light stress.
transition) is potentially more susceptible to photodamage (Finazzi et al., 2001), so that the core phosphorylation could also be effective to protect cores after the dissociation of LHCII upon state transition.

Alternatively, core subunit phosphorylation might be specifically required for undocking of the peripheral antenna. Our results revealed that the strongest phosphorylation of PSII subunits was found in Fraction III (Figure 7). It is thus possible that the simultaneous phosphorylation of both minor LHCIs and PSII core subunits facilitates dissociation of most LHCIs from the PSII core complex. The crystal structure of the PSII core complex (dark green) is from 2AXT.pdb, and the LHCII monomer (light blue) and trimer (light green) structures are from 2NHW.pdb. Structural organizations of the megacomplex and supercomplex refer to Dekker and Boekema (2005) and Nield et al. (2004), respectively. The first- and second-step phosphorylations are indicated by blue and red circles with a letter “P,” respectively.

(4) LHCII Type III (LhcbM2 and -7) and Type IV (LhcbM1)

In contrast with LHCII type I, LHCII type III (LhcbM2 and -7) and IV (LhcbM1) were not associated with PSII when they were phosphorylated (Figures 7 and 8). These unbound phosphorylated polypeptides were not only in the State 2 samples but also in the State 1 samples. These results may imply that the phosphorylation-induced undocking of LHCII type II occurs irrespective of the state and plays a unique role in photosynthesis other than state transitions. Care should be taken, however, since the quantity of phosphorylated polypeptides cannot be directly compared between State 1 and 2 gels, as approximately the same amount of proteins were resolved in the two 2-DE gels although the amounts of unbound LHCIs in State 1 cells should be significantly lower than those in State 2 cells (Figures 8C and 8D). It is unclear whether LHCII type III and IV are more phosphorylated under State 2. Simply put, their behavior upon phosphorylation is different from that of LHCII type I; they leave PSII upon phosphorylation.
It should be noted that a mutant of the *LhcBM1* gene in *C. reinhardtii* (*npq5*) has been shown to undergo normal state transitions (Elrad et al., 2002). Interestingly, the *npq5* mutant was originally isolated as exhibiting little energy-dependent quenching (*qE* quenching) (Niyogi et al., 1997). This implies that the phosphorylated (dissociated) LhcbM1 and -2 may, in fact, be involved in *qE* quenching in addition to their roles in state transitions. Considering that *C. reinhardtii* displays little to no expression of the essential protein for *qE* quenching, PsbS (Niyogi et al., 2005), and only has a limited capacity of *qE* quenching when grown under normal light (Finazzi et al., 2006), the underlying mechanism for *qE* quenching in this green alga is not necessarily the same as that elucidated in higher plants (see reviews in Horton and Ruban, 2005; Niyogi et al., 2005). Further investigation on the significance of phosphorylation of LhcbM1 and -2 will clarify this point.

Based on the above observations, we propose the following model for the State 1 to 2 transition in *C. reinhardtii*: (1) unphosphorylated LHCII megacomplexes are localized in the appressed region (State 1); (2) the phosphorylation of LHCII type I in the major LHCII trimer triggers the division of the megacomplex, resulting in individual C2S2 supercomplexes; (3) the phosphorylation of CP26 and CP29 as well as of PSI core subunits D2 and CP43 induces the undocking of all LHCII megacomplexes from PSI; and (4) free LHCII megacomplexes reassociate with the PSI-LHCI supercomplex yielding State 2 (illustrated in Figure 10A).

**Ultrastructure of Thylakoid Membranes in State Transitions**

It has been inferred that state transitions and the ultrastructure of thylakoid membranes are closely related (Allen and Forsberg, 2001). The functional absorption cross section of PSI is reported to be large in grana and small in stroma lamellae (Garab and Mustárdy, 1999; Danielsson et al., 2006; Veerman et al., 2007), indicating that PSI in the stacked region of the thylakoid membrane tends to harbor more antenna proteins. In fact, a highly ordered paracrystalline arrangement of supercomplexes, from which the megacomplexes are thought to be solubilized, was observed only in the highly stacked grana core in *Arabidopsis* (Boekema et al., 2000). In contrast with the grana/stroma lamellae-type structure in higher plants, thylakoid membranes in *Chlamydomonas* are organized such that long bands of two to four thylakoid membranes are extensively but loosely appressed (Bertos and Gibbs, 1998). If the megacomplexes described above are localized in the appressed region, the thylakoid membranes in *C. reinhardtii* need to have a larger appressed region in State 1 than in State 2 to accommodate the larger number of megacomplexes. Indeed, an electron micrograph of *C. reinhardtii* thin sections from State 1 cells show a higher degree of membrane stacking, whereas those in State 2 were relatively unstacked (see Supplemental Figure 6 online). Currently, it is unclear whether state transitions affect the ultrastructure of thylakoid membranes or vice versa.

Figure 10B shows a model of state transitions that incorporates stacking/unstacking of thylakoid membranes. Since PSI megacomplexes are localized in the appressed region (Boekema et al., 2000), the diffusion of LHCII proteins is restricted (Kirchhoff et al., 2004) under State 1 conditions. After the membranes unstack, which may be caused by the phosphorylation of LHCII type I, more LHCII proteins, including CP26, CP29, and the core proteins CP43 and D2, are phosphorylated, leading to the undocking of all peripheral antenna proteins from PSI, which is partly reflected in the phosphorylation kinetics (Figure 9).

Further studies will be needed to characterize the significance of our other findings, including the phosphorylation of LHCII type III and IV in State 1, the redox-dependent phosphorylation of PSI core subunits, and possible changes in the thylakoid ultrastructure during state transitions. Such studies will provide new perspectives on the mutually related mechanisms of photosynthetic acclimation, including state transitions, *qE* quenching, and repair of photodamaged PSI centers.

**METHODS**

**Strain and Growth Conditions**

A genetically modified *Chlamydomonas reinhardtii* strain PsbB-His, carrying a 6x His-tag at the C terminus of the *psbB* gene (Suzuki et al., 2003), was grown in Tris-acetate-phosphate medium (Gorman and Levine, 1965) under a light intensity of 20 μmol photons m⁻² s⁻¹ aerated with ambient air at 23°C.

**Induction of State Transitions**

PsbB-His cells were treated as described previously (Bülté et al., 1990) to lock State 1 and 2. Briefly, the cells were incubated with 10 μM DCMU or 5 μM FCCP for 20 min under the same conditions as their growth to induce State 1 or State 2, respectively.

**Isolation of Thylakoid Membranes and PSII Particles**

Cells were harvested at mid-log phase (5 × 10⁶ cells/mL), and thylakoid membranes were isolated similar to methods described by Chua and Bennoun (1975) with minor modifications. The buffer used for preparing thylakoid membranes contained 25 mM MES, 0.33 M sucrose, and 1.5 mM NaCl, pH 6.5. Disruption of cells was done using a BioNeb system (Glas-Col) at 10 kgf/cm². Thylakoid membranes were adjusted to 1 mg chlorophyll/mL (for isolating PSI-LHCII supercomplexes) in a buffer containing 25 mM MES, 100 mM NaCl, and 12.5% (w/v) glycero, pH 6.5, and solubilized with 1.0 to 1.2% (w/v) TM (Anatrace) at 4°C for 30 min in the dark. Solubilized thylakoid membranes were applied to a column with ProBond Resin (Invitrogen), which was preequilibrated with the same buffer supplemented with 15 mM imidazole and 0.03% (w/v) TM. Chromatography was performed as described previously (Suzuki et al., 2003). The eluate was diluted 10 times with a buffer containing 25 mM MES, pH 6.5, and 0.03% TM and concentrated to 1 mg chlorophyll/mL using an Amicon Ultra 100 KMWCO filter. Chlorophyll concentrations were determined as described by Porra et al. (1989) unless otherwise indicated.

**Gel Filtration**

PSII complexes isolated by nickel affinity chromatography (20 μg chlorophyll) were separated on a SMART System with a Superdex 200 PC 3.2/30 column (GE Healthcare), which was preequilibrated with 25 mM MES and 0.03% TM, pH 6.5. Gel filtration was conducted at 10°C with a flow rate of 25 μL/min. Protein concentrations were detected from their absorbance at 280 nm. Molecular masses of the protein complexes were estimated with a standard curve drawn with blue dextran (molecular mass of 2000 kD), thyroglobulin (molecular mass of 669 kD), and apoferritin (molecular mass of 443 kD) from GE Healthcare.

---

**Strain and Growth Conditions**

A genetically modified *Chlamydomonas reinhardtii* strain PsbB-His, carrying a 6x His-tag at the C terminus of the *psbB* gene (Suzuki et al., 2003), was grown in Tris-acetate-phosphate medium (Gorman and Levine, 1965) under a light intensity of 20 μmol photons m⁻² s⁻¹ aerated with ambient air at 23°C.

**Induction of State Transitions**

PsbB-His cells were treated as described previously (Bülté et al., 1990) to lock State 1 and 2. Briefly, the cells were incubated with 10 μM DCMU or 5 μM FCCP for 20 min under the same conditions as their growth to induce State 1 or State 2, respectively.

**Isolation of Thylakoid Membranes and PSII Particles**

Cells were harvested at mid-log phase (5 × 10⁶ cells/mL), and thylakoid membranes were isolated similar to methods described by Chua and Bennoun (1975) with minor modifications. The buffer used for preparing thylakoid membranes contained 25 mM MES, 0.33 M sucrose, and 1.5 mM NaCl, pH 6.5. Disruption of cells was done using a BioNeb system (Glas-Col) at 10 kgf/cm². Thylakoid membranes were adjusted to 1 mg chlorophyll/mL (for isolating PSI-LHCII supercomplexes) in a buffer containing 25 mM MES, 100 mM NaCl, and 12.5% (w/v) glycero, pH 6.5, and solubilized with 1.0 to 1.2% (w/v) TM (Anatrace) at 4°C for 30 min in the dark. Solubilized thylakoid membranes were applied to a column with ProBond Resin (Invitrogen), which was preequilibrated with the same buffer supplemented with 15 mM imidazole and 0.03% (w/v) TM. Chromatography was performed as described previously (Suzuki et al., 2003). The eluate was diluted 10 times with a buffer containing 25 mM MES, pH 6.5, and 0.03% TM and concentrated to 1 mg chlorophyll/mL using an Amicon Ultra 100 KMWCO filter. Chlorophyll concentrations were determined as described by Porra et al. (1989) unless otherwise indicated.

**Gel Filtration**

PSII complexes isolated by nickel affinity chromatography (20 μg chlorophyll) were separated on a SMART System with a Superdex 200 PC 3.2/30 column (GE Healthcare), which was preequilibrated with 25 mM MES and 0.03% TM, pH 6.5. Gel filtration was conducted at 10°C with a flow rate of 25 μL/min. Protein concentrations were detected from their absorbance at 280 nm. Molecular masses of the protein complexes were estimated with a standard curve drawn with blue dextran (molecular mass of 2000 kD), thyroglobulin (molecular mass of 669 kD), and apoferritin (molecular mass of 443 kD) from GE Healthcare.
Polypeptides were separated by SDS-PAGE as described previously (Ikeuchi and Inoue, 1988). The first dimension of 2-DE, isoelectric focusing, was performed according to the procedure described by Hippler et al. (2001) with an Immobiline DryStrip (pH 4 to 7) on a Multiphor II system (GE Healthcare). The second dimension, SDS-PAGE, was conducted as described above. Polypeptide bands were stained with Quick CBB (Wako) or silver stain MS kit (Wako).

Immunoblotting was performed as described previously (Takahashi et al., 2006). Antibodies against D1, D2, CP43, CP47, PsbO, PsbP, and PsbQ (1/5000 dilution) were kind gifts from M. Ikeuchi at the University of Tokyo, those against LhcbM6 for detecting LHCII type I and P25K for detecting LHCII type II and IV (1/10,000 dilution) were provided by M. Hippler at the University of Münster, and those against CP26, CP29, LhcbM5 (1/10,000 dilution), and Psaa/B (1/5000 dilution) were described by Takahashi et al. (2006). Phosphorylated proteins were detected with an antiphosphothreonine (42H4) monoclonal antibody (1/5000 dilution) from New England Biolabs. Anti-rabbit secondary antibody was used for all but antiphosphothreonine monoclonal antibody for which anti-mouse secondary antibody was used. To identify phosphorylated polypeptides, the duplicated blots were checked either with antibodies specific to each polypeptide (SDS-PAGE) or MS/MS analysis (2-DE). Quantification of protein levels was done with ImageJ software (Abramoff et al., 2004).

In-gel trypsin digestion of polypeptides was performed as described by Shevchenko et al. (1996). The digested samples were applied to a Paradigm MS4 dual solvent system (Michrom BioResources). Solutions of 2 and 90% (v/v) acetonitrile with 0.1% (v/v) formic acid were used as the mobile phases A and B, respectively. The flow rate was 2 μL/min with a gradient as follows: 5% of B from 0 to 40 min, 95% from 40 to 50 min, and 5% from 50 to 60 min. Mass spectra were measured with a Finningan LTQ linear ion trap mass spectrometer (Thermo Fisher Scientific). Obtained mass spectra were analyzed by SEQUEST software (revision 3.3; Thermo Fisher Scientific) using a genomic database for C. reinhardtii (version 3.0 assembly). The obtained peptide information was filtered based on the following criteria: number of top matches = 1, probability < 0.001, and Sf-final score > 0.85.

**Fluorescence Analysis**

Steady state fluorescence spectra at 77K were measured as described previously (Takahashi et al., 2006). Functional absorption cross section of PSII (F_PsaA) was determined according to Kolber et al. (1998) with a fluorescence induction and relaxation (FIRE) fluorimeter (Satlantic). Samples were dark-adapted for 10 min, and a 100-μs single-turnover flash was applied by a blue LED (455 nm) to cumulatively saturate PSII. The absolute F_PsaA values were calculated according to the calibration factor provided by the manufacturer.

**Phosphorylation Kinetics from State 1 to State 2**

PsbB-His cells grown as described above were illuminated with far-red light through a band-pass filter (720 ± 10 nm) for 15 min to induce State 1, which was followed by 10 min illumination with blue light through a band-pass filter (467 ± 10 nm) to induce State 2. Cells were sampled every 2 min in the course of transition to State 2. Polypeptides were isolated from the SDS-solubilized cells by precipitating with chloroform/methanol (Wessel and Flugge, 1984) and subjected to SDS-PAGE. Phosphorylation of polypeptides was detected by immunoblotting using an antiphosphothreonine antibody as described above. Three independent experiments were performed.

**HPLC Analysis**

Pigments of protein samples were extracted with 80% (v/v) acetone. HPLC was performed using a 4.6 × 250 mm VP-ODS C18 column (Shimadzu). Separation of pigments was done by a linear gradient of acetone/water as described previously (Niyogi et al., 1997). Molar extinction coefficients of 36,620 and 122,510 M⁻¹ cm⁻¹ were used for neoxanthin and β-carotene, respectively.

**Transmission Electron Microscopy**

Observation of single-particle PSII complexes was done according to Ortega et al. (2000). An H-7650 ZeroA electron microscope was used, and micrographs were digitally captured at a magnification of ×30,000. For observation of thylakoid membranes, cells induced to State 1 or State 2 were fixed in 1% glutaraldehyde in 0.1 M cacodylate buffer, pH 7.4, and 1% osmium tetroxide, dehydrated, and embedded in epoxy resin. Ultrathin sections were stained with 2% uranyl acetate and 0.2% lead citrate. A JEM 1200EX electron microscope (JEOL) was used, and samples were imaged at a magnification of ×50,000.

**Supplemental Data**

The following materials are available in the online version of this article.

- **Supplemental Figure 1.** Specificity of Nickel Affinity Chromatography.
- **Supplemental Figure 2.** Polypeptide Composition in Gel Filtration Fractions I to III during States 1 and 2.
- **Supplemental Figure 3.** Relative Abundance of the PSII Core Subunits in the Gel Filtration Fractions from State 1– and State 2-Locked Samples.
- **Supplemental Figure 4.** Distribution of PSII Particle Projections on Electron Micrographs.
- **Supplemental Figure 5.** Evidence for the Correspondence between Spot #12 in Figure 8D and CP29.
- **Supplemental Figure 6.** Structure of Thylakoid Membranes in State 1 and State 2.
- **Supplemental Table 1.** Relative Protein Levels of the PSII Core Subunits in the Three PSII Fractions.
- **Supplemental Table 2.** MS/MS Analysis of the LHCII Proteins Bound to the PSII Complexes.
- **Supplemental Table 3.** MS/MS Analysis of the Free LHCII Proteins in the Nickel Affinity Chromatography Flow-Through.

**ACKNOWLEDGMENTS**

We thank Wesley Swingley for critical comments on the manuscript. We thank Shin-ichiro Ozawa at Okayama University for the help with MS/MS. We also thank Masahiko Ikeuchi and Michael Hippler for the kind gift of the antibodies. Our work was supported by the Research Fellowship for Young Scientists from the Japan Society for the Promotion of Science (1804526 to M.I.), Grants-in-Aid for Scientific Research for Plant Graduate Student from the Nara Institute of Science and Technology (to M.I.), and Grants-in-Aid for Scientific Research from the Ministry of Education, Culture, Sports, Science, and Technology (18GS0318 to Y.T. and J.M.).

Received March 11, 2008; revised August 4, 2008; accepted August 7, 2008; published August 29, 2008.

**REFERENCES**


Molecular Remodeling of Photosystem II during State Transitions in *Chlamydomonas reinhardtii*

Masakazu Iwai, Yuichiro Takahashi and Jun Minagawa

*Plant Cell*; originally published online August 29, 2008;
DOI 10.1105/tpc.108.059352

This information is current as of January 18, 2018

<table>
<thead>
<tr>
<th>Supplemental Data</th>
<th>/content/suppl/2008/08/19/tpc.108.059352.DC1.html</th>
</tr>
</thead>
<tbody>
<tr>
<td>eTOCs</td>
<td>Sign up for eTOCs at: <a href="http://www.plantcell.org/cgi/alerts/ctmain">http://www.plantcell.org/cgi/alerts/ctmain</a></td>
</tr>
<tr>
<td>CiteTrack Alerts</td>
<td>Sign up for CiteTrack Alerts at: <a href="http://www.plantcell.org/cgi/alerts/ctmain">http://www.plantcell.org/cgi/alerts/ctmain</a></td>
</tr>
<tr>
<td>Subscription Information</td>
<td>Subscription Information for <em>The Plant Cell</em> and <em>Plant Physiology</em> is available at: <a href="http://www.aspb.org/publications/subscriptions.cfm">http://www.aspb.org/publications/subscriptions.cfm</a></td>
</tr>
</tbody>
</table>